



Composition and diversity of culturable cyanobacteria in sediment samples from the upper layers of two tropical reservoirs

Composición y diversidad de cianobacterias cultivables en muestras de sedimentos superiores de dos embalses tropicales

Lina Arismendi-González^{1*}, Esteban Arroyave², Luz Yaneth Orozco¹, Clara María Arboleda-Baena³, Marisol Sepúlveda-Sánchez¹, Hilda Palacio-Betancur⁴, Carlos Enrique Muskus-López⁵, Elisabeth Pohlen⁶, María Teresa Flórez-Molina¹, Jaime Palacio-Baena¹

Abstract

Benthic cyanobacteria research in high mountain reservoirs remains limited, mainly due to their complexity and knowledge gaps that persist in relation to their ecology in tropical regions. This study aimed to explore the composition, diversity, and toxic potential of cyanobacteria in the upper sediment of two Colombian reservoirs. Our investigation involved multiple methodologies, such as germination experiments, that allowed us to assess the presence and viability of cyanobacteria in upper sediment samples, while the competitive ELISA assay allowed for the quantification of toxins within the cultures. The molecular analysis of Operational Taxonomic Units (OTUs) from subsamples of sediment cultures focused on evaluating cyanobacterial diversity and richness among prokaryotic phyla, and the Phylogenetic analysis of culturable cyanobacteria. Chlorophyll-*a* measurements confirmed the presence of viable populations in sediment cultures, while microscopic identification demonstrated the growth capacity of cyanobacteria from the orders Nostocales, Chroococcales, Oscillatoriales, and Synechococcales under controlled laboratory conditions. Despite low microcystin levels in culture, the prior detection of *mcy* genes in direct sediment samples suggests a possible toxic potential of cyanobacteria inhabiting the upper sediments. Community analysis, based on the OTUs abundance, revealed a notably diverse microbial community in both reservoir sediments, with a higher relative abundance of cyanobacteria compared to other prokaryotic phyla. These findings support the hypothesis that surface sediments play a fundamental role as a repository for cyanobacteria that may pose inherent risks to ecosystem health. In conclusion, this research underscores the necessity of further studies to achieve a holistic comprehension of benthic cyanobacteria dynamics in high-mountain tropical reservoirs.

Keywords: benthic zones, cyanobacterial diversity, sediment, tropical reservoir

Resumen

Las investigaciones sobre cianobacterias bentónicas en reservorios de alta montaña en regiones tropicales aún son limitadas, debido a su complejidad y vacíos de conocimiento en relación con su ecología. Este estudio tuvo como objetivo explorar la composición, diversidad y potencial tóxico de estas cianobacterias en sedimentos superiores de dos embalses colombianos, empleando múltiples

¹ Grupo de Gestión y Modelación Ambiental (GAIA), Universidad de Antioquia, Medellín, Colombia.

² University of Texas Medical Branch (UTMB), Galveston, Texas, USA.

³ Laboratory of Microbial Processes & Biodiversity, Departamento de Hidrobiología, Universidade Federal, São Carlos, Brazil.

⁴ Universidad CES, Medellín, Colombia.

⁵ Programa de Estudio y Control de Enfermedades Tropicales (PECET), Universidad de Antioquia, Medellín, Colombia.

⁶ Institute for Animal Ecology, and Systematics, Justus Liebig University, Gießen, Germany.

* Corresponding author: lina.arismendi@udea.edu.co

Received: October 12, 2023; accepted: November 8, 2023; published: February 6, 2024.

metodologías, como experimentos de germinación para evaluar su presencia y viabilidad en sedimentos, ensayos de ELISA competitivo para cuantificar toxinas en cultivo, análisis de Unidades Taxonómicas Operativas (OTUs) a partir de submuestras de los cultivos para evaluar su diversidad y riqueza entre los diferentes phylum procarióticos, y análisis filogenético de cianobacterias cultivables. Se confirmó la presencia de poblaciones viables de cianobacterias en los sedimentos mediante mediciones de clorofila-*a* y la identificación microscópica demostró su capacidad de crecimiento en condiciones controladas de laboratorio. A pesar de los bajos niveles de microcistina en cultivo, la detección previa de genes *mcy* en muestras directas de sedimento, sugiere un potencial tóxico en las cianobacterias presentes. El análisis de la comunidad reveló una notable diversidad microbiana en los sedimentos de ambos embalses, con una mayor abundancia relativa de cianobacterias en comparación con otros phylum procarióticos. Estos hallazgos respaldan la hipótesis de que los sedimentos superficiales desempeñan un papel fundamental como depósito de cianobacterias que pueden plantear riesgos inherentes a la salud del ecosistema. En conclusión, esta investigación subraya la necesidad de realizar más estudios para lograr una comprensión holística de la dinámica de las cianobacterias bentónicas en reservorios tropicales de alta montaña.

Palabras clave: diversidad de cianobacterias, embalse tropical, sedimento, zona bentónica

INTRODUCTION

Cyanobacteria are a group of photosynthetic prokaryotes that are among the oldest and most successful forms of life, with a diverse range of functional traits that allow them to colonize various environments and cope with adverse conditions (Dalu and Wasserman, 2018; Roque et al., 2021). In tropical regions, permanently high temperatures in aquatic ecosystems have been demonstrated to promote the dominance of this phylum of phytoplankton over other forms throughout the year (Chorus and Bartram, 1999; Dalu and Wasserman, 2018; Harke et al., 2016; Paerl et al., 2011; Reynolds and Petersen, 2000).

Several studies suggest that surface sediments act as reservoirs for cyanobacteria in aquatic ecosystems and indicate that the benthic zone may be an important source of cyanotoxins, even in the absence of planktonic surface cyanobacteria (Bormans et al., 2020; Gangi et al., 2020; Izaguirre et al., 2007; Westrick and Szlag, 2018). Furthermore, benthic environmental samples collected in warmer regions have been found to contain significantly more toxins than those collected in temperate climates (Gaget et al., 2022), underscoring the necessity of advancing our understanding of their ecology to comprehend their dynamics in tropical freshwater systems (Quiblier et al., 2013).

Most investigations on benthic cyanobacteria in tropical and subtropical regions have been conducted in shallow environments (Borges et al., 2015; Crispino and Sant'anna, 2006; Magina and

da Silva e Silva, 2008; Noffke et al., 2003; Silva-e-Silva, 2004), and the knowledge of these organisms in high mountain reservoirs is still limited. The overall perspective suggests that research on benthic cyanobacteria is lagging, and the risk associated with them remains largely unknown (Gaget et al., 2022); Colombian reservoirs are not far from this problem.

Although the increase in eutrophication and water temperature favors the proliferation of cyanobacteria in tropical aquatic ecosystems, there is no systematic record of cyanobacterial blooms or a protocol for their control and management in Colombia (Salomón et al., 2020). Current knowledge about the taxonomy and ecology of cyanobacteria in Colombian reservoirs is mainly limited to taxonomic lists of planktonic species, while the ecological characteristics and toxic potential of cyanobacteria from the benthic zone are unknown.

These knowledge gaps were highlighted by Quiblier et al. (2013) and more recently by Gaget et al. (2022) in their respective studies, each proposing that one reason for this is that water utilities rarely establish monitoring practices for benthic cyanobacteria or associated secondary metabolites.

The present study aimed to elucidate the role of surface sediments as a reservoir of cyanobacterial with toxic potential in two tropical reservoirs. For this, we conducted an initial germination experiment with sediments samples and evaluated their viability and toxic potential, as described in

Arismendi-González et al. (2021). Subsequently, we performed molecular analyses on the cultured samples to gain insights into the richness, diversity, and composition of the cyanobacterial community in the surface sediments of both reservoirs.

These analyses were carried out under laboratory conditions, and our main objective was to establish a baseline for future research on the ecology and potential risks associated with benthic cyanobacteria in these systems. Our findings provide valuable information on the presence and characteristics of cyanobacteria in two tropical reservoirs and may aid in the development of strategies for their management and control.

MATERIALS AND METHODS

Study area

Riogrande II reservoir is located northwest of Medellín (Antioquia), Colombia (75°32'30", 75°26'10"W and 6°33'50", 6°28'07" N) with a maximum flood level of 2270 m.a.s.l. It is mainly

used for drinking water withdrawal, but also for electric power generation (up to 321MW). This system has a surface area of 12.14 km², with a total volume of 240 million m³, and a maximum depth of 47.2 m. The water temperature span ranges from 15 °C to 25 °C and the mean annual rainfall of the area is approximately 2250 mm (Figure 1a).

Porce II reservoir is located northwest of Antioquia, Colombia (75°09'14", 75°04'59" W and 6°44'57", 6°48'45"N) with a maximum flood level of 925 m.a.s.l. It is mainly used for electric power generation (405MW). The reservoir has a surface area of 9.8 km², with a mean volume of 231 million m³, a maximum depth of 96.5 m and the mean annual rainfall of the area is approximately 1960 mm (Figure 1b).

Sampling

From December 2014 to September 2015, sediment samples were collected at three stations in each reservoir (Figure 1). A UWITEC gravity nucleator equipped with transparent acrylic

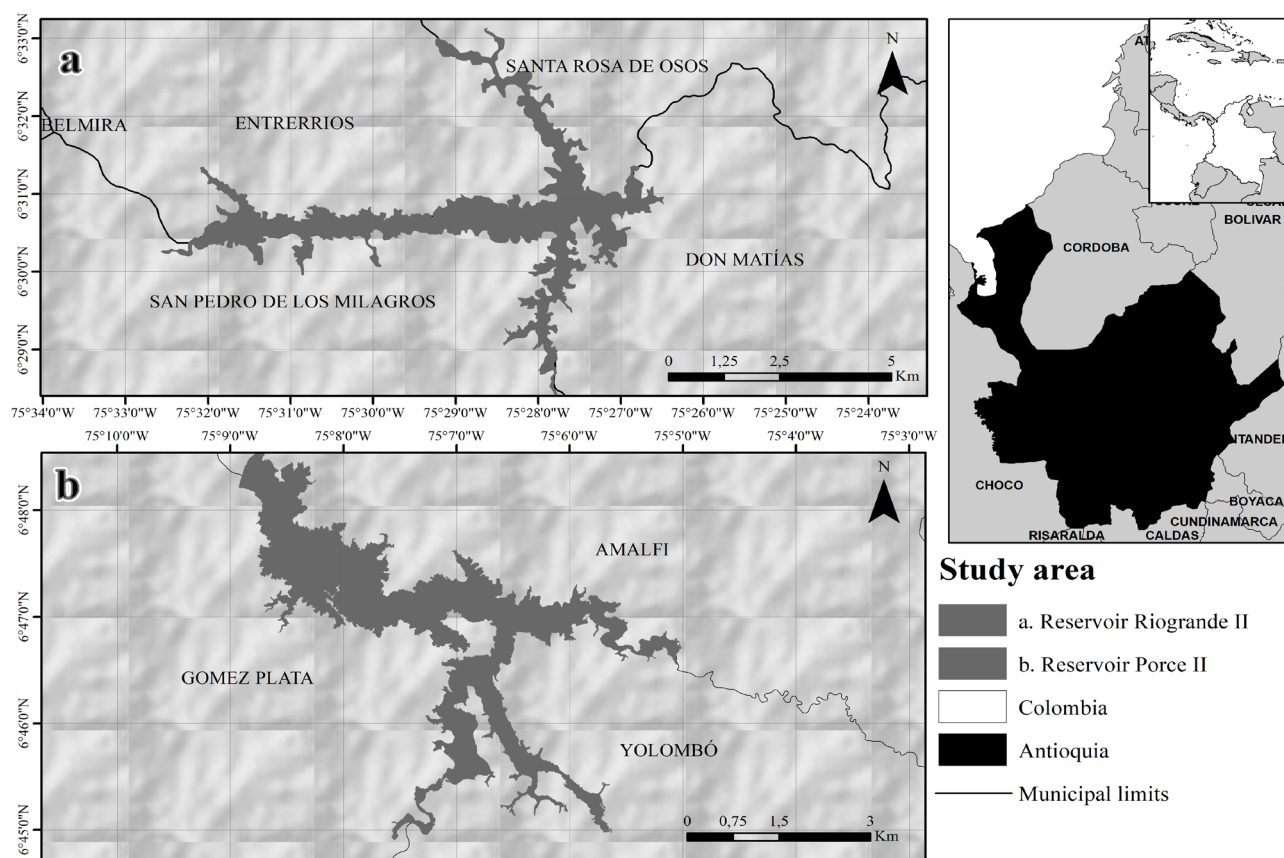


Figure 1. Location of study areas, Riogrande II and Porce II reservoirs.

tubes, approximately 50 cm in length and 6 cm in diameter, was employed for sediment sampling collection. All samples were kept at 4 °C. Triplicate samples were obtained from each station of the same reservoir at all four sampling events.

Laboratory analysis

Cyanobacterial cultures from sediment samples

Cyanobacterial cultures were established from environmental samples collected from the oxic and silty upper layer sediments (0-4 cm). Five g of sediment samples were aseptically transferred into sterile 250 mL glass bottles containing 50 mL of ASM-1 medium. Cultures were incubated at 25 °C in a climate chamber using dark/light (1300 lx) cycles of 12 h, as previously described by Arismendi-González et al. (2021). The cultures were monitored every 20 days. However, the evaluation of the cultures was conducted at the end of the experiment, 300 days later.

Identification of cultured cyanobacteria from sediment samples

To confirm the presence and viability of cyanobacteria in the cultures, chlorophyll-*a* measurements were performed using the FluoroProbe III Workstation 25 (bbe-Moldaenke, Kiel, Germany) following the methodology outlined in Arismendi-González et al. (2021). This allowed us to differentiate each phytoplankton group and verify cyanobacteria growth in each culture sample. To identify cyanobacteria, we used an Olympus CX31 microscope at 400X magnification. Additionally, random samples from the successfully isolated cultures were analyzed using a JSM-6490LV Scanning Electron Microscope-SEM (Jeol, USA). Identification was performed following the Komárek and Anagnostidis (1998; 2005), Komárek (2013), and Tucci et al. (2019) taxonomic keys.

Quantification of Microcystins by ELISA in culture from sediment samples

Microcystins (MCs) production by cyanobacterial cultures was assessed by enzyme-linked immunosorbent assay (ELISA) with the Envirologix QuantiPlate commercial kit (Envirologix Inc., Portland, ME, USA). The ELISA assay had a limit of detection between 0.16 to 2.5 ppb. Before analysis, all samples were subjected to three

freeze/thaw cycles to disrupt the cellular matrix and promote toxin release. The samples were further processed by sonication using a Branson model 2510R-DTH Ultrasonic Cleaner (Branson Ultrasonic Corp., USA) to facilitate cell lysis and toxin release, then ELISA test was performed following the manufacturer's instructions.

Molecular analysis of cyanobacterial diversity in cultures from sediment samples

Sub-samples from each reservoir (viable cultures from all sampling stations were included) were pooled and prepared for DNA extraction. Total genomic DNA was extracted from each sub-sample using a genomic DNA mini kit (Invitrogen™, K1820-02) according to manufacturers' recommendations, and DNA concentration was assessed using Quant-it PicoGreen dsDNA Assay (Invitrogen™, USA).

Amplification and sequencing of 16S rRNA gene by Next Generation Sequencing-NGS

The 16S rRNA gene was full-length amplified (1500pb) using the primers 27F1 (5' AGAGTTTGATCCTGGCTCAG 3') y 1492R (5' TACGGYTACCTTGTTACGACTT 3') (Heuer et al., 1997) and the quality of the amplicons was assessed by 1% agarose gel electrophoresis. The samples were sequenced by the Illumina MiSeq platform generating paired reads of 300pb. The V3-V4 hypervariable regions from the 16S rRNA gene were amplified by PCR using the universal bacterial primers Bakt_341F (5' CCTACGGGNGGCWGCAG 3') and Bakt_805R (5' GACTACHVGGGTATCTAATCC 3') (Herlemann et al., 2011), both with Illumina adapter, pad, and linker sequence.

16S amplicon sequence analyses

Sequences were analyzed using the Mothur software (Schloss et al., 2009) according to Miseq Standard Operating Procedure. The quality control analysis of the sequencing data was performed through the FastQC program, selecting only the sequences with a Q value > 30 and the VSEARCH program was applied to identify and remove the chimeric sequences. The Operational Taxonomic Units (OTUs) were defined with a minimum of 97% identity by taxonomy assignment using the RDP reference database sequences as a guide. In

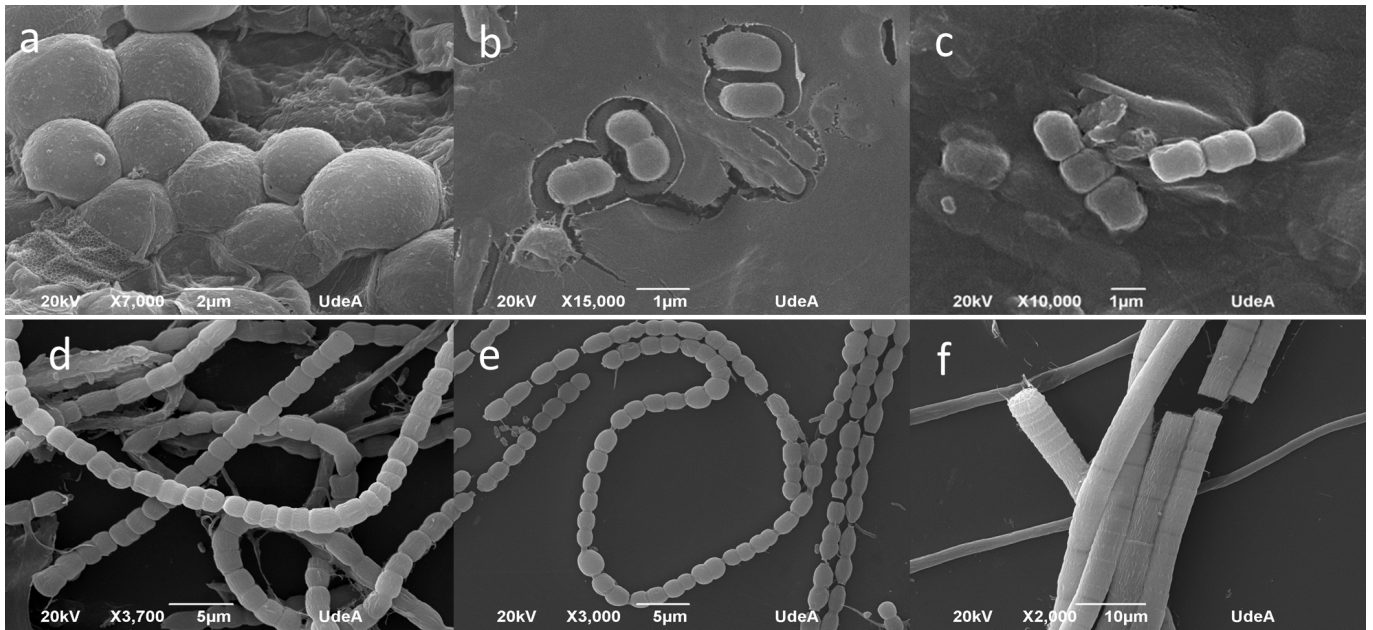


Figure 2. Cyanobacterial order observed by Scanning Electron Microscope from cultures of sediment samples a-b) Chroococcales, c) Synechococcales, d-e) Nostocales, f) Oscillatoriales.

with values close to the detection limit (0.23 and 0.27 µg/L), while in Porce II microcystin was detected in cultures from the three stations, with concentrations of MCs that ranged between 0.16 and 2.5 µg/L (Table S1).

Genetic diversity of cyanobacteria in culture from sediment samples

Community analyses of OTUs sequences

Bioinformatic analysis using the Mothur package yielded 34,530 reads in Porce II and 20,479 reads in Riogrande II, with coverage value of 0.961 and 0.963, respectively (Table 2).

The Shannon index from both reservoirs' samples showed that Riogrande II exhibited a higher diversity compared to Porce II. Furthermore, Riogrande II demonstrated a greater uniformity of species, as indicated by a Shannon evenness

value of 0.70. In contrast, the Chao index revealed that Porce II had the highest richness of microbial communities among the two reservoirs.

The dominant phyla in the upper sediment microbial community of Porce II and Riogrande II reservoirs were Cyanobacteria, Proteobacteria, and Planctomycetes, which accounted for almost three-quarters of all reads. It should be noted that the relative abundance of Cyanobacteria was high in both reservoirs, compared to the total microbial community (Figure 3).

The diversity and richness of cyanobacteria were evaluated based on the abundance of Operational Taxonomic Units (OTUs) belonging to the Cyanobacteria phylum. Out of the total 3629 OTUs acquired, 671 were identified as cyanobacteria through the NCBI database. In addition, to uphold the integrity of our final analysis, we exclusively considered sequences that demonstrated an

Table 2. Microbial community diversity indices in sediment cultures from Porce II and Riogrande II

Samples	Number of Reads	Coverage	Number of OTUs (richness)	Simpson index	Shannon index	Chao index
PII	34,530	0,961	2183	24,715	4,718	6328,114
RGII	20,479	0,963	1446	46,746	5,119	3029,757

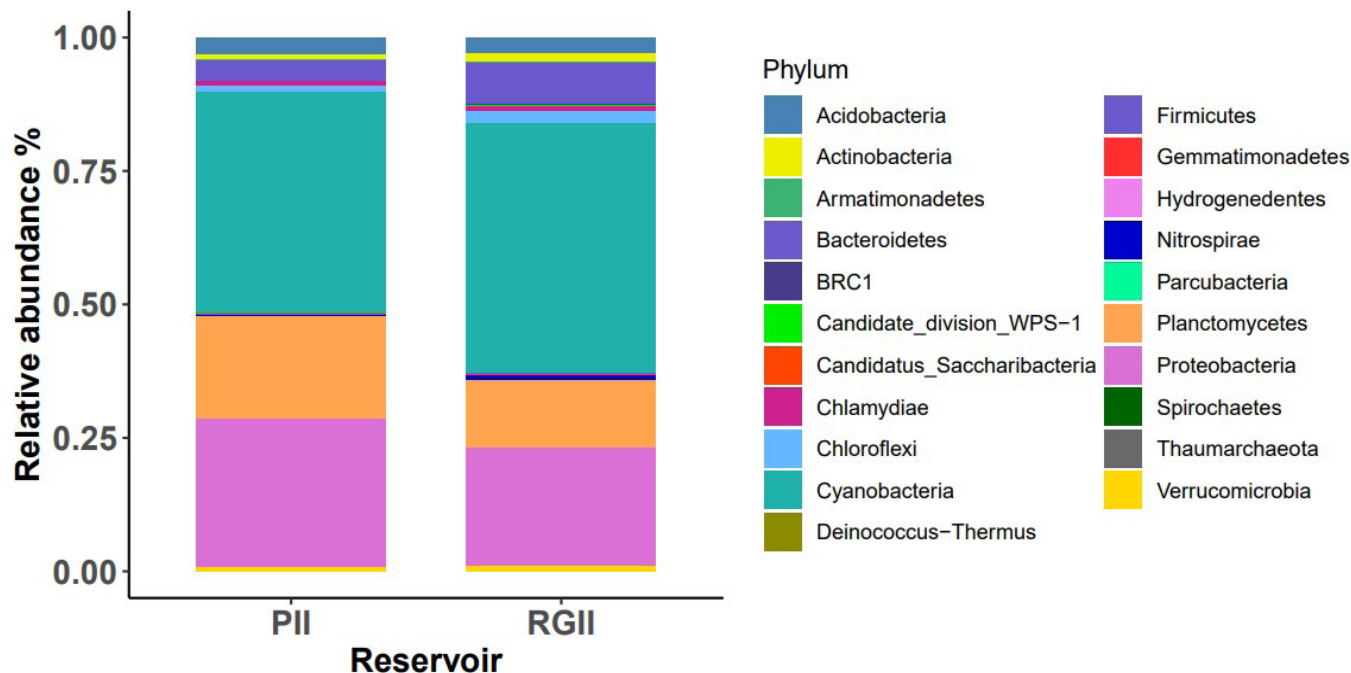


Figure 3. Relative abundance of different prokaryotic phyla (Bacteria and Archaea) in sediment cultures from Porce II and Riogrande II reservoirs.

identity percentage of at least 95%.

As seen in Figure 4, the cyanobacterial community diversity was similar for both reservoirs. However, cyanobacterial OTUs richness was higher in Porce II reservoir. Of the 342 OTUs reported for Porce II, 22 were found to be shared with Riogrande II, representing the genera *Anabaena*, *Arthronema*, *Calothrix*, *Cyanobacterium*, *Desmonostoc*, *Leptolyngbya*, *Nostoc*, *Pantalinema*, *Phormidesmis*, *Phormidium*, *Pseudanabaena*, *Roholtiella*, and *Tolypothrix*.

According to Table S2, the highest number of

OTUs in both reservoirs belonged to the orders Nostocales and Synechococcales. However, the most dominant OTUs were assigned to the genus *Cyanobacterium* (87) in Porce II and *Leptolyngbya* (42) in Riogrande II.

Phylogenetic analysis of culturable cyanobacteria strains

The phylogenetic analysis indicated that the cyanobacterial communities in these reservoirs are represented in four orders (Nostocales, Chroococcales, Oscillatoriales, and Synechococcales), as observed through the

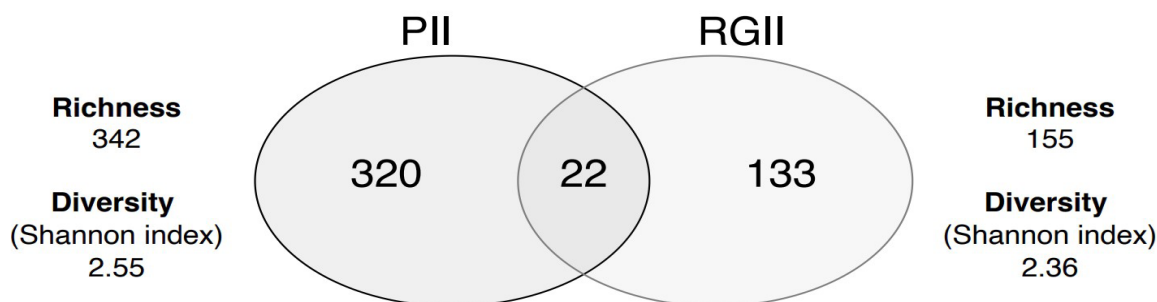


Figure 4. Venn diagram showing the number of shared and unique cyanobacterial OTUs in the Porce II and Riogrande II reservoirs, as well as their diversity and richness of cyanobacteria as a function of OTUs abundance.



that their growth patterns are not homogeneous within the culture containers (Quiblier et al., 2013) which could lead to erroneous estimates of the true abundance of the community, our results show how some communities of cyanobacteria settled down at the bottom of the reservoirs, survived, and were still viable, as reported by previous studies by Everson et al. (2011) and Gangi et al. (2020). Moreover, spectral analyses of the cultures showed that the mean concentration of chlorophyll-*a* corresponding to the cyanobacterial communities was higher in Riogrande II, which was consistent with a higher species richness observed for this reservoir through the microscopic analysis of cultures.

Despite the quantification of microcystins in cultures from upper sediment samples in both reservoirs was low, we consider that it constitutes a warning about the possible toxic potential of these populations, a hypothesis that is supported by the detection and quantification of *mcy* genes in direct upper sediment samples of both reservoirs as shown in Arismendi-González et al. (2021), and in other results not yet published (Table S3). Additionally, as shown in Bormans et al. (2020), the sole presence of *mcy* genes in sediment samples suggests that the populations present may pose a risk to organisms due to their toxic potential and their resuspension potential in the water column. However, these results lead us to consider the possibility that cyanobacterial populations present in the sediments of these reservoirs could be producing other types of toxins not included in this study, which is why we suggest a broader toxin screening be considered in future research.

According to the analysis of OTUs sequences, the upper sediment samples from both reservoirs showed a high diversity of microbial communities. However, the sample from Porce II reservoir exhibited the highest species richness. On the other hand, the relative abundance of cyanobacteria was high with respect to the whole community found in culture sediment samples from both reservoirs. This finding is significant as it strengthens the hypothesis suggesting that the surface sediment constitutes an important sink for cyanobacteria, as indicated in a study conducted by Wood et al. (2021), who propose that the presence of cyanobacterial 16S rRNA genes in sediment cores provides strong evidence for cyanobacterial sedimentation in reservoirs. Furthermore, the

coexistence of benthic and planktonic strains within the clades suggests that germinated cyanobacteria may belong to both habitats.

Although many of the taxa detected through phylogenetic analysis have planktonic habits, they do not totally coincide with the planktonic taxa previously described for these ecosystems (Arboleda-Baena, 2017; Palacio et al., 2015; Rodríguez, 2013; Sepúlveda-Sánchez et al., 2021). Similar results are described in the study by Wood et al. (2021) who found that none of the strains from their study were very closely related to the planktonic as well as toxin-producing taxa isolated from the Baltic Sea and various lakes in Finland, previously reported by Lyra et al. (2001) and Rajaniemi et al. (2005).

Finally, the microscopic and molecular analysis coincided on the presence of cyanobacteria of the orders Nostocales, Chroococcales, Oscillatoriales, and Synechococcales in the sediments of both reservoirs, with a greater predominance of Nostocales, which may be related to the high number of akinetes that was observed in some culture samples during germination experiments. Similar results are described in the studies of Hellweger et al. (2008) and Legrand et al. (2017a). These findings are relevant as they may indicate past blooms and contribute to knowledge about the formation of future blooms, as several studies have reported that these structures could persist and germinate from aquatic sediment samples (Gangi et al., 2020; Legrand et al., 2017b; Legrand et al., 2019; Wood et al., 2009; Wood et al., 2021) after several years. In addition, this is the first time that *Fisherella* and *Hapalosiphon* have been reported in the Porce II reservoir.

This was the first study to investigate benthic archives of cyanobacteria in the upper sediments of two Colombian reservoirs, allowing us to demonstrate through different methods the presence of active populations that reflect how these sediments provide a favorable environment for their development and possibly allow them to maintain their activity over time, which could have implications not only related to the quality of the water and the ecology of the reservoir for their possible implications in the formation of superficial blooms but also their possible repercussions on Public Health. However, this topic has not yet been explored in depth, which is why we recommend

developing new research to confirm whether sediment cores can function as a proxy to predict possible blooms of planktonic cyanobacteria in reservoirs. Finally, this research highlighted the importance of further studies to achieve a holistic understanding of benthic cyanobacteria dynamics in high-mountain tropical reservoirs.

ACKNOWLEDGMENTS

This study was supported by COLCIENCIAS (658 - 2014, project 111565843084) and Empresas Públicas de Medellín (EPM). We would like to thank all the staff at the GAIA group, Universidad de Antioquia-UdeA, and the staff at EPM for the logistical support for the monitoring of the reservoir.

AUTHOR CONTRIBUTION

Lina Arismendi-Gonzalez: conceptualization, methodology, investigation, writing-original draft, writing-review and editing. Esteban Arroyave: methodology, formal analysis, writing-original draft, writing-review and editing. Luz Yaneth Orozco: methodology, formal analysis, writing-review and editing. Clara Arboleda-Baena: methodology, investigation, formal analysis, writing-review and editing. Marisol Sepulveda-Sanchez: methodology, investigation, formal analysis, writing-review and editing. Hilda Palacio-Betancur: formal analysis, writing-review and editing. Carlos Muskus-Lopez: conceptualization, writing-review and editing, supervision. Elisabeth Pohlson: writing-review and editing, supervision. Maria Flórez Molina: conceptualization, writing-review and editing, supervision, project administration. Jaime Palacio Baena: conceptualization, writing-review and editing, supervision, project administration.

CONFLICT OF INTEREST

The authors declare that they have no financial/commercial conflicts of interest.

REFERENCES

Arboleda Baena, C. M. (2017). *Determinación del potencial tóxico de cianobacterias Nostocales y Chroococcales en la columna de agua del embalse Riogrande II (Antioquia) a través de la detección del gen mcy* [Tesis de Maestría, Universidad de Antioquia]. <https://catalejo.udea.edu.co/cgi-bin/koha/opac-detail.pl?biblionumber=768343>

- Arismendi-González, L., Sepúlveda-Sánchez, M., Arboleda-Baena, C. M., Palacio-Betancur, H., Murillo-Ramos, E., Muskus-López, C. E., Pohlson, E., Flórez-Molina, M. T., Betancur-Uran, J., & Palacio-Baena, J. (2021). Evidence for toxic cyanobacteria in sediments and the water-sediment interface of a tropical drinking water reservoir. *Limnologica*, 91, 125924. <https://doi.org/10.1016/j.limno.2021.125924>
- Borges, H. L., Branco, L. H., Martins, M. D., Lima, C. S., Barbosa, P. T., Lira, G. A., Bittencourt-Oliveira, M. C., & Molica, R. J. (2015). Cyanotoxin production and phylogeny of benthic cyanobacterial strains isolated from the northeast of Brazil. *Harmful Algae*, 43, 46-57. <https://doi.org/10.1016/j.hal.2015.01.003>
- Bormans, M., Savar, V., Legrand, B., Mineaud, E., Robert, E., Lance, E., & Amzil, Z. (2020). Cyanobacteria and cyanotoxins in estuarine water and sediment. *Aquatic Ecology*, 54(2), 625-640. <https://doi.org/10.1007/s10452-020-09764-y>
- Chorus, I., & Bartram, J. (1999). *Toxic Cyanobacteria in Water: a guide to their Public Health consequences, monitoring, and management* [Archivo PDF]. <https://cdn.who.int/media/docs/default-source/wash-documents/water-safety-and-quality/toxic-cyanobacteria---1st-ed.pdf>
- Crispino, L. M., & Sant'Anna, C. L. (2006). Cianobacterias marinhas bentônicas de ilhas costeiras do Estado de Sao Paulo, Brasil. *Revista Brasileira de Botânica*, 29(4), 639-656. <https://doi.org/10.1590/S0100-84042006000400014>
- Dalu, T., & Wasserman, R. J. (2018). Cyanobacteria dynamics in a small tropical reservoir: Understanding spatio-temporal variability and influence of environmental variables. *Science of the Total Environment*, 643, 835-841. <https://doi.org/10.1016/j.scitotenv.2018.06.256>
- Everson, S., Fabbro, L., Kinnear, S., & Wright, P. (2011). Extreme differences in akinete, heterocyte and cylindrospermopsin concentrations with depth in a successive bloom involving *Aphanizomenon ovalisporum* (Forti) and *Cylindrospermopsis raciborskii* (Woloszynska) Seenaya and Subba Raju. *Harmful Algae*, 10(3), 265-276. <https://doi.org/10.1016/j.hal.2010.10.006>
- Gaget, V., Almuhtar, H., Kibuye, F., Hobson, P., Zamyadi, A., Wert, E., & Brookes, J. D. (2022). Benthic cyanobacteria: A utility-centred field study. *Harmful Algae*, 113, 102185. <https://doi.org/10.1016/j.hal.2022.102185>
- Gangi, D., Plastani, M. S., Laprida, C., Lami, A., Dubois, N., Bordet, F., Gogorza, C., Frau, D., & de Tezanos Pinto, P. (2020). Recent cyanobacteria abundance in a large sub-tropical reservoir inferred from analysis of sediment cores. *Journal of Paleolimnology*, 63(3), 195-209. <https://doi.org/10.1007/s10933-020-00110-8>
- Harke, M. J., Steffen, M. M., Gobler, C. J., Otten, T.

- G., Wilhelm, S. W., Wood, S. A., & Paerl, H. W. (2016). A review of the global ecology, genomics, and biogeography of the toxic cyanobacterium, *Microcystis* spp. *Harmful Algae*, 54, 4-20. <https://doi.org/10.1016/j.hal.2015.12.007>
- Heuer, H., Krsek, M., Baker, P., Smalla, K., & Wellington, E. M. (1997). Analysis of actinomycete communities by specific amplification of genes encoding 16S rRNA and gel-electrophoretic separation in denaturing gradients. *Applied Environmental Microbiology*, 63(8), 3233-3241. <https://doi.org/10.1128/aem.63.8.3233-3241.1997>
- Hellweger, F. L., Kravchuk, E. S., Novotny, V., & Gladyshev, M. I. (2008). Agent-based modeling of the complex life cycle of a cyanobacterium (*Anabaena*) in a shallow reservoir. *Limnology and Oceanography*, 53(4), 1227-1241. <https://doi.org/10.4319/lo.2008.53.4.1227>
- Herlemann, D. P., Labrenz, M., Jürgens, K., Bertilsson, S., Waniek, J. J., & Andersson, A. F. (2011). Transitions in bacterial communities along the 2000 km salinity gradient of the Baltic Sea. *The ISME Journal*, 5, 1571-1579. <https://doi.org/10.1038/ismej.2011.41>
- Izaguirre, G., Jungblut, A. D., & Neilan, B. A. (2007). Benthic cyanobacteria (Oscillatoriaceae) that produce microcystin-LR, isolated from four reservoirs in southern California. *Water Research*, 41(2), 492-498. <https://doi.org/10.1016/j.watres.2006.10.012>
- Komárek, J., & Anagnostidis K. (1998). *Cyanoprokaryota. Teil 1/part 1: Chroococcales*. Springer Spektrum, Heidelberg.
- Komárek, J., & Anagnostidis K. (2005). *Cyanoprokaryota. Teil 2/Part 2: Oscillatoriales*. Springer Spektrum.
- Komárek, J. (2013). *Cyanoprokaryota. Teil 3/part 3: Heterocytous Genera*. Springer Spektrum.
- Legrand, B., Le Jeune, A. H., Colombet, J., Thouvenot, A., & Latour, D. (2017a). Akinetes may be representative of past nostocalean blooms: A case study of their benthic spatiotemporal distribution and potential for germination in a eutrophic lake. *Applied and Environmental Microbiology*, 83(23), 01571-17. <https://doi.org/10.1128/AEM.01571-17>
- Legrand, B., Lamarque, A., Sabart, M., & Latour, D. (2017b). Benthic archives reveal recurrence and dominance of toxigenic cyanobacteria in a eutrophic lake over the last 220 years. *Toxins*, 9(9), 271. <https://doi.org/10.3390/toxins9090271>
- Legrand, B., Miras, Y., Beauger, A., Dussauze, M., & Latour, D. (2019). Akinetes and ancient DNA reveal toxic cyanobacterial recurrences and their potential for resurrection in a 6700-year-old core from a eutrophic lake. *Science of the Total Environment*, 687, 1369-1380. <https://doi.org/10.1016/j.scitotenv.2019.07.100>
- Lyra, C., Suomalainen, S., Gugger, M., Vezie, C., Sundman, P., Paulin, L., & Sivonen, K. (2001). Molecular characterization of planktic cyanobacteria of *Anabaena*, *Aphanizomenon*, *Microcystis*, and *Planktothrix* genera. *International Journal of Systematic and Evolutionary Microbiology*, 51, 513-526. <https://10.1099/00207713-51-2-513>
- Magina, F., & da Silva e Silva, L. (2008). Cianobacterias Psamicas em Sedimentos Marginais da Lagoa Rodrigo de Freitas, Estado do Rio de Janeiro, Brasil. *Anuario Do Instituto de Geociencias*, 31, 43-49.
- Novfke, N., Gerdes, G., & Klenke, T. (2003). Benthic cyanobacteria and their influence on the sedimentary dynamics of peritidal depositional systems (siliciclastic, evaporitic salty, and evaporitic carbonatic). *Earth-Science Reviews*, 62(1-2), 163-176. [https://doi.org/10.1016/S0012-8252\(02\)00158-7](https://doi.org/10.1016/S0012-8252(02)00158-7)
- Paerl, H. W., Hall, N. S., & Calandrino, E. S. (2011). Controlling harmful cyanobacterial blooms in a world experiencing anthropogenic and climatic-induced change. *Science of the Total Environment*, 409(10), 1739-1745. <https://doi.org/10.1016/j.scitotenv.2011.02.001>
- Palacio, H. M., Ramírez, J. J., Echenique, R. O., Palacio, J. A., & Sant'anna, C. L. (2015). Floristic composition of cyanobacteria in a neotropical, eutrophic reservoir. *Revista Brasileira de Botânica*, 38(4), 865-876. <https://doi.org/10.1007/s40415-015-0185-3>
- Palacio, J., Florez-Molina, M., Molina-Perez, F., Toro, M., Gomez, A., Penuela, G., & Cuellar, W. (2017). *Estudio de la Problemática Ambiental de tres Embalses de Empresas Públicas de Medellín para la Gestión Integral y Adecuada del Recurso Hídrico* [Informe convenio EPM- Universidad de Antioquia].
- Quiblier, C., Wood, S., Echenique-Subiabre, I., Heath, M., Villeneuve, A., & Humbert, J. F., (2013). A review of current knowledge on toxic benthic freshwater cyanobacteria ecology, toxin production and risk management. *Water Research*, 47(15), 5464-5479. <https://doi.org/10.1016/j.watres.2013.06.042>
- Rajaniemi, P., Hrouzek, P., Kaštovská, K., Willame, R., Rantala, A., Hoffmann, L., Komárek, J., & Sivonen, K. (2005). Phylogenetic and morphological evaluation of the genera *Anabaena*, *Aphanizomenon*, *Trichormus* and *Nostoc* (Nostocales, cyanobacteria). *International Journal of Systematic and Evolutionary Microbiology*, 55(1), 11-26. <https://doi.org/10.1099/ij.s.0.63276-0>
- Reynolds, C. S., & Petersen, A. C. (2000). The distribution of planktonic Cyanobacteria in Irish lakes in relation to their trophic states. *Hidrobiología*, 424, 91-99. <https://doi.org/10.1023/A:1003901012233>
- Rodríguez López, M. L. (2013). *Cultivo, aislamiento e identificación de cianobacterias procedentes de los embalses la Fe, Riogrande II y Porce II* [Tesis de Maestría, Universidad de Antioquia].
- Roque J., & Tamagnini, P. (2021). *Characterization of*



- cyanobacterial strains isolated from Portuguese soil/soil biocrusts and a first insight into this community* [Tese de Mestrado, Universidade do Porto]. <https://repositorio-aberto.up.pt/bitstream/10216/139400/2/528160.pdf>
- Salomón, S., Rivera-Rondón, C. A., & Zapata, A. M. (2020). Cyanobacterial blooms in Colombia: State of knowledge and research needs in the context of climate global change. *Revista de La Academia Colombiana de Ciencias Exactas, Físicas y Naturales*, 44(171), 376-391. <https://doi.org/10.18257/raccefyn.1050>
- Schloss, P. D., Westcott, S. L., Ryabin, T., Hall, J. R., Hartmann, M., Hollister, E. B., Lesniewski, R. A., Oakley, B. B., Parks, D. H., Robinson, C. J., Sahl, J. W., Stres, B., Thallinger, G. G., Van Horn, D. J., & Weber, C. F. (2009). Introducing Mothur: open-source, platform independent, community-supported software for describing and comparing microbial communities. *Applied and Environmental Microbiology*, 75(23), 7537-7541 <https://doi.org/10.1128/AEM.01541-09>.
- Sepúlveda-Sánchez, M., Arismendi-González, L. M., Arboleda-Baena, C. M., Muskus-López, C. E., Pohlen, E., Florez-Molina, M. T., & Palacio-Baena, J. A. (2021). First evidence of potential toxic cyanobacteria in the water-sediment interface of a tropical drinking water reservoir. *Revista Internacional de contaminación Ambiental*, 37, 259-272. <https://doi.org/10.20937/RICA.53696>
- Silva-e-Silva, L., (2004). Estruturas microbianas recentes da lagoa Pernambuco, Estado do Rio de Janeiro, Brasil. *Revista Brasileira de Paleontologia*, 7(2), 189-192. <https://doi.org/10.4072/rbp.2004.2.11>.
- Tucci, A., Sant'Anna, C. L., Azevedo, M. T. P., Malone, C. F. S., Werner, V. R., Rosini, E. F., Gama, W. A., Hentschke, G. S., Osti, J. A. S., Dias, A. S. Jacinavicius, F. R., & Santos, K. R. S. (2019). *Atlas de Cianobactérias e Microalgas de Águas Continentais Brasileiras* [Arquivo PDF]. https://www.infraestruturameioambiente.sp.gov.br/institutodebotanica/wp-content/uploads/sites/235/2013/09/virtuais_3atlas.pdf
- Westrick, J. A., & Szlag, D., (2018). A cyanotoxin primer for drinking water professionals. *American Water Works Association*, 110(8), E1-E16. <https://doi.org/10.1002/awwa.1088>
- Wood, S. A., Jentsch, K., Rueckert, A., Hamilton, D. P., & Cary, S. C. (2009). Hindcasting cyanobacterial communities in Lake Okaro with germination experiments and genetic analyses. *FEMS Microbiology Ecology*, 67(2), 252-260. <https://doi.org/10.1111/j.1574-6941.2008.00630.x>
- Wood, S. M., Kremp, A., Savelle, H., Akter, S., Vartti, V. P., Saarni, S., & Suikkanen, S. (2021). Cyanobacterial Akinete Distribution, Viability, and Cyanotoxin Records in Sediment Archives from the Northern Baltic Sea. *Frontiers in Microbiology*, 12, 681881. <https://doi.org/10.3389/fmicb.2021.681881>