



## Diversity of cave Phlebotomines (Diptera: Psychodidae) from a Colombian cave

Manuela Velásquez Londoño<sup>a</sup>, Adam M.M. Stuckert<sup>b</sup>, Rafael J. Vivero<sup>a,c</sup>, Daniel R. Matute<sup>b,\*</sup>

<sup>a</sup> Programa de Estudio y Control de Enfermedades Tropicales, Universidad de Antioquia

<sup>b</sup> Biology Department, University of North Carolina, 250 Bell Tower Drive, Genome Sciences Building, Chapel Hill, NC 27510, United States

<sup>c</sup> Grupo de Microbiodiversidad y Bioprospección, Universidad Nacional de Colombia, Sede Medellín

### ARTICLE INFO

#### Keywords:

Sandflies  
Leishmaniasis  
Vector biology  
Cave biodiversity  
Species richness

### ABSTRACT

Sandflies are vector species of *Leishmania*, among many other pathogens, with a global distribution and a variety of ecological niches. Previous samplings have found that karstic formations (i.e., caves, grottos, and folds formed by the erosion of limestone) serve as a natural habitat to sandfly species. The majority of samplings of cave sandfly diversity have occurred in Brazil and to date none have studied the species composition in a cave in the Northern Andes. We collected sandflies in the Cave “Los Guácharos”, in the state of Antioquia, Colombia. The sampling was carried out during two consecutive nights in September 2019. CDC-type light traps were installed inside the cavern and in other surrounding karst systems (caves, rock-breaks, and folds). In total, we identified 17 species of sandfly from the cave and surrounding karst systems, including a new record for Colombia (*Bichromomyia olmeca*), and provide the first karstic reports for four other species (*Lutzomyia gomezi*, *Lutzomyia hartmanni*, *Pintomyia ovallesi*, and *Psychodopygus panamensis*). We then used the results of our survey and published literature to test two hypotheses. First, that sandfly diversity in Neotropical caves is richest nearer to the equator, and second that there is a phylogenetic signal of karstic habitat use in sandflies. Counter to our predictions, we found no evidence that diversity follows a latitudinal gradient. Further, we find no evidence of a phylogenetic signal of karstic habitat use, instead finding that the use of caves likely evolved multiple times across several genera. Our results highlight the importance of a wide sampling to understand the natural habitat of sandflies and other disease vectors.

### 1. Introduction

The family Psychodidae is a speciose group of dipterans that is globally distributed. The family encompasses seven different subfamilies and has over 4,000 species (Pave et al., 2011; Bejarano and Estrada 2016). The majority of the species diversity is encompassed by two subfamilies. The subfamily Psychodinae (Wagner 2004; Espíndola et al., 2012) commonly referred to as moth flies or drain flies, which includes 2,000 species, many of which are human commensals (Cordeiro and Wagner 2018; Munstermann 2019). The second subfamily, Phlebotominae, includes 500 species, of which over 90 are vectors of diseases of humans and other animals alike (Killick-Kendrick 1990; Ready 2013). These species are commonly referred to as sandflies and are the main vectors of Leishmaniasis in the tropics and subtropics. Just for Leishmaniasis, one of the diseases transmitted by members of the subfamily Phlebotominae, more than 12 million people are infected and over 2

million new cases are reported annually. The number of recorded deaths due to *Leishmania* infection per year is around 60,000, a number that is, in all likelihood, a vast underestimate due to inadequate reporting requirements and *Leishmania* prevalence in areas with minimal access to healthcare (Alvar et al., 2012; Okwor and Uzonna 2016; Karimkhani et al., 2016; Bailey et al., 2017). Sandflies also transmit other pathogens that cause severe diseases, among which are three arboviruses (Sicilian virus, Naples virus, and Toscana virus; (Tesh et al., 1975; Batieha et al., 2000; Dionisio et al., 2003; Izri et al., 2008)), *Vesiculovirus* (Comer and Tesh 1991), *Orbivirus* (Depaquit et al., 2010; Phumee et al., 2021), *Flavivirus* (Acevedo et al., 2008), and *Bartonella bacilliformis*, the etiological agent of Carrion's disease (Caceres et al., 1997).

Species from the Psychodidae family occupy a variety of habitats. Psychodinae species are often associated with synanthropic habitats but are also commonly found in drains and aquatic environments (Cordeiro and Wagner 2018). Phlebotominae sandflies are often associated with

\* Corresponding author.

E-mail address: [dmatute@email.unc.edu](mailto:dmatute@email.unc.edu) (D.R. Matute).

<https://doi.org/10.1016/j.actatropica.2022.106515>

Received 8 January 2022; Received in revised form 12 May 2022; Accepted 12 May 2022

Available online 20 May 2022

0001-706X/© 2022 The Authors. Published by Elsevier B.V. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

places where they can obtain blood meals such as human settlements (Ramos et al., 2014) and coffee plantations (Ferro et al., 2015). Mating often occurs at lekking sites at the base of large trees (Memmott 1991, 1992). Females tend to oviposit close to these lekking sites in clay rich soils. Painstaking collections have revealed the presence of immature stages in such sites (Rutledge and Mosser 1972; Vivero et al., 2015). The specificity of habitat varies across species, and in some cases even populations, and while some species are geographically widespread and inhabit a variety of habitats others seem to be more restricted in their geographic and ecological distribution. The precise details of habitat usage by most species remain unknown (Alexander 2000).

One of the potential ecological niches of sandflies is karstic landscapes (Oca-Aguilar et al., 2013; Blavier et al., 2019; Costa et al., 2021). Karstic landscapes are irregular limestone formations affected by erosion that encompass fissures, sinkholes, underground streams, and caverns. Previous reports have identified that some Phlebotomine species frequently breed in cavern systems. Quate (1962) carried out the first cave collection in the Batu Caves (Malaysia) and identified 22 phlebotomids from different subfamilies. Since then, it has become clear that cavern soil is a potential breeding ground for Psychodids, especially for Phlebotomines (e.g., Quate 1962; Galati et al., 2003; Alves et al., 2011; Campos et al., 2017; Costa et al., 2021). Several species are exclusively found in caves and breeding in caves is a trait that is widespread in the family (Alves et al. 2008; Carvalho et al. 2010; Carvalho et al., 2011; Barata et al., 2012; Vilela et al., 2015). Cave surveys have the potential to add crucial information about the taxonomic and ecological diversity of sandflies.

South and Central America harbor a high diversity of Phlebotomine species (D'Agostino et al., 2022) and they house the vector genus *Lutzomyia sensu lato*. Samplings from Brazil suggest the existence of species restricted to caves (Alves et al., 2011; Carvalho et al., 2013; Campos et al., 2017), and that important disease vectors can breed in caves (Cutolo et al., 2008; Carvalho et al., 2017). Few efforts have addressed the diversity of this group in caves in the Neotropics, especially in Northern South America. The only study on this matter found Phlebotomines associated with a karstic, not cavernous, system in the Cañón del Río Claro Natural reserve (Antioquia, Colombia). Bejarano et al. (2018) reported the presence of *Warileya (Hertigia) hertigi*, a Phlebotomine not involved in the transmission of disease. Addressing the diversity of Phlebotomines in caves in the Northern Andes holds the potential to reveal general patterns about the diversity of this vector group, which remains a largely unaddressed question.

We expanded the previous sampling of Bejarano et al. (2018) and collected Phlebotomines in the cave system in the Los Guácharos Cave in the Cañón del Río Claro Natural reserve (Antioquia, Colombia). Our point sampling revealed a species previously unknown to occur in Colombia, and four species that were not known to occur in karstic environments. We then used our findings as well as existing datasets to test the hypothesis that Neotropical cave sandfly species diversity follows the expectations of a latitudinal species richness gradient. We find that cave sandfly species diversity does not follow the expectations of a latitudinal species richness gradient and that the species diversity in tropics and subtropics seems to be similar. Finally, we used comparative phylogenetic methods to study whether association to karstic environments has evolved multiple times in the evolutionary history of the Psychodidae family. We find that association to karst is not a trait with a strong phylogenetic signal. Our findings suggest that the ecological and evolutionary implications of cave diversity is fundamental for the understanding of species richness in the family.

## 2. Methods

### 2.1. Locality

The focus of our collection was the cave system at the Cañón del Río Claro Nature Reserve. The reserve is located between the municipalities

of Sonsón, Puerto Triunfo and San Luis, to the southwest of the department of Antioquia (5° 5' N, 74° 39' W, (Vivero et al., 2010). This cave system is of karstic origin and was formed by the erosion of irregular limestone (Restrepo Martínez 2011). The largest cave of the cave system, the Los Guácharos cave, has a length of 442.8 m, with an entrance 10 to 15 m high by 2 to 3 m wide and is considered a humid cave (Moncada et al., 1989). The environment inside the karstic formation can be classified as caverns, fissures, sinkholes, and underground streams. The annual mean temperature and mean relative humidity of the cave are 22 °C and 80% respectively. The cave serves as a roosting site for oilbirds (*Steatornis caripensis*) and chestnut long-tongued bats (*Lionycteris spurrelli*, Caballero et al. 2021).

### 2.2. Sampling

To characterize the entomofauna in the cave, we carried out a sampling during two consecutive nights in the month of September 2019. We installed 10 CDC-type light traps for a period of 12 h starting at 18:00. The traps were collected at 6:00 the following morning. To attract blood feeding insects, we placed dry ice under the traps. The sublimating CO<sub>2</sub> mimics the respiration of vertebrates and increases the sandfly yield. Two of the CDC traps were located in a cave with a 2 m<sup>2</sup> cavity. Two more traps were placed inside a karst fold with a depth of 4 m and an altitude of approximately 4 m. The other six traps were placed in the main gallery of the cavern. The location of the traps obeyed safety precautions and the depth of bodies of water (Fig. 1). For comparison, we installed a CDC trap in the vegetation outside the cave. We compared the species richness outside and inside the cave.

For species identification, we transported the nets of the CDC traps to the Laboratory of Medical Entomology of the Program for the Study and Control of Tropical Diseases of the University of Antioquia. We extracted specimens from the traps using an aspirator and placed them in a Petri dish and inspected under a Leica microscope. To clear samples, we immersed each sandfly in lactophenol (1:1 ratio). Finally, we identified samples to species-level following sandfly taxonomic classification keys (Galati 2019, Young and Duncan 1994, Ibañez-Bernal 1999).

### 2.3. Species diversity and comparison to other studies

Next, we calculated metrics of species diversity in the Los Guácharos cave and compared them to other cave collections in the Neotropics to determine whether sandfly species richness followed a latitudinal gradient. We only included studies for which the raw data was reported (i.e., all the species collected during the sampling and their respective abundances), we compiled data from 32 other cave samplings. Using this dataset, we calculated three different metrics of species diversity. First, we used the number of collected species (i.e., species richness). Second, we used the Shannon's Index ( $H$ ) which is a weighted geometric mean of species abundances.  $H$  follows the form:

$$H = -\sum_{i=1}^R [p_i \times \ln(p_i)]$$

where  $p_i$  is the proportional representation of a given species and  $R$  is the number of species in the sample. If most of the abundance in the sample is concentrated in a single species, then  $H$  will be 0. In cases in which all the species are equally abundant,  $H$  will equal  $\ln(R)$ .

We calculated a third index of diversity, species evenness, which is defined as  $H$  normalized by the number of species and follows the form:

$$J = H / \log(\text{species number})$$

$J$  ranges from 0 (i.e., low evenness, presence of a dominant species) and 1 (i.e., complete evenness, all species represented equally). All metrics were calculated using the R package *vegan* (function *diversity*, (Oksanen 2013; Oksanen et al., 2013, 2015)).



Fig. 1. CDC light traps in the Los Guácharos cave collection. A-B. Cave. C. Vegetation outside the cave. D. Karstic folds.

Different samplings had different levels of effort. To account for differences in sampling effort, we used a rarefaction approach in which we estimated the expected number of species in each locale with the smallest number of collected samples from a published cave sampling study which we have included in our expanded dataset ( $N = 6$ ). We also estimated rarefaction curves for all the 33 localities. For these analyses we used the functions *rarefy* in the R package *vegan* and used 1000 bootstrap replicates per locality (Oksanen 2013; Oksanen et al., 2013, 2015). This function calculates a rarefaction curve and estimates total species abundance for a given sample size (in our case  $N = 6$ ), thus correcting for sampling effort across studies.

We next studied whether there was a relationship between different metrics of cave sandfly species richness (both before and after rarefaction) and latitude. Our expectation was that species richness in caves would follow a latitudinal pattern of diversity and that caves closer to the tropics would show higher species richness, as has been observed for other taxa (Mittelbach et al., 2007; Mannion et al., 2014; Jablonski et al., 2017; Pontarp et al., 2019). We used linear regressions to assess whether the different metrics of species richness were associated with the latitude of the cave (function *lm*, library *stats*; R Core Team 2021). We bootstrapped the regression coefficients using the function *Boot* ( $R = 1,000$ ; library *car*, Fox et al., 2012. Fox and Weisberg 2019).

#### 2.4. The evolution of cavernicolous niche

Our work and others report several unrelated species in the Psychodidae family to have a cavernicolous niche. This suggests the possibility that the niche has evolved several times in the evolutionary history of the group. We formally addressed whether cavernicolous niche has evolved more than once in the evolutionary history of the family using comparative phylogenetic methods. We used a previously published phylogeny of the group using four different loci (D'Agostino et al., 2022) and classified each species as cavernicolous (i.e., has been collected in caves) or non-cavernicolous. We measured whether cave habitat followed a phylogenetic pattern. We used Pagel's  $\lambda$  (Pagel 1999) for discrete traits (i.e., whether a species had been found in caves or not). Pagel's  $\lambda$ , is a measure of phylogenetic signal which estimates the extent to which the phylogenetic history of a clade is predictive of the trait distribution at the tree tips. Values of  $\lambda$  lower than 1.0 represent traits being less similar amongst species than expected from their phylogenetic relationships. A  $\lambda$  equal to 1.0 suggests that traits covary with phylogeny (Pagel 1997, 1999) and is consistent with niche conservatism along the phylogeny (Cooper et al., 2010). A value close to zero indicates no effect of the phylogenetic history on the evolution of a trait. We used the wrapper *fitLambda* for the *ace* function (library *ape*, Paradis et al., 2004; Paradis and Schliep 2019) for ancestral character estimation and  $\lambda$  calculation. This function optimizes Pagel's  $\lambda$  tree transformation for a

discrete character evolving by a continuous-time Markov chain. We also used the function *fitdiscrete* (library *geiger*, (Harmon et al., 2008; Pennell et al., 2014)) which yielded the same results.

### 3. Results

#### 3.1. Collection description

We collected 52 sandflies, 16 males and 36 females. This difference in the sex ratio of the collection obeys different patterns of attraction to light traps (Toprak and Özer 2007). The samples belonged to 17 species of the Phlebotominae subfamily, representatives of the genera *Warileya*, *Lutzomyia*, *Micropygomyia*, *Bichromomyia*, *Evandromyia*, *Pintomyia*, *Pressatia*, *Psathyromyia*, *Pschoathyrdomyia*, *Pschoathyromygia*, and *Helcorcitomyia* (Table 1). The most abundant species was *Warileya (Hertigia) hertigi* (21.14%), followed by *Pintomyia ovallesi* (17.3% of total samples), and *Lutzomyia gomezi* (11.53%; Table 1). This collection represents two novel findings. First, one of the collected species (*Bichromomyia olmeca nociva*) is novel for Colombia which significantly expands the range of the species. Second, we report four species that had not been, or have been rarely, collected in caves.

#### 3.2. New report of *Bichromomyia olmeca nociva* for Colombia

Our sampling includes one species that had not been reported for Colombia. We describe the morphological features of this specimen as follows.

##### 3.2.1. *Bichromomyia olmeca nociva*

(Young and Arias 1982): folding (5° 53' 16.8729 "N, 74 ° 51' 17.6029" W), Cañón del Río Claro Nature Reserve (Antioquia); Fig. 2A-C. (♂), interocular distance (1.5 mm) equivalent to 1/3 of the width of the eye (0.5 mm), epandrial lobe (2.3 mm) shorter than the gonocoxite (2.5 mm), ratio of aedeagal ducts (3.0 mm) / sperm pump (1.3 mm) equals to 3.0:1.0. *Bichromomyia olmeca nociva* was found in the inner part of the cave across the river. Before this report, *B. olmeca nociva* has been reported for the Brazilian Amazon but not for the Colombian territory (~1,700 km extension).

#### 3.3. New reports for cave-dwelling species

Our sampling found four species that have not been collected in caves, and one species which has only been reported in a cave once

before. In this sampling, we found that *Lutzomyia hartmanni* (Fairchild and Hertig 1957) appeared associated with two different karstic environments, caves and rock-breaks. *Lutzomyia gomezi* (Nitzulescu 1931) has not been reported before in association with caves or any karstic systems. However, in this report we find the species in the Los Guácharos cave and surrounding vegetation. In both instances we found females that recently had a blood meal. Finally we observed two more species, *Psychodopygus panamensis* (Shannon 1926) and *Pintomyia ovallesi* (Ortiz 1951) in caves for the first time in our sampling. *Psychodopygus panamensis* is a generalist species that is often characterized as anthropophilic (González et al., 1999; Santamaría et al., 2020; Rigg et al., 2021) as is *Pintomyia ovallesi* (Felicciangeli 1997; Felicciangeli and Rabinovich 1998; Rabinovich and Felicciangeli 2004). One previous study has collected samples from this species in a cave in Belize (Williams 1976). In both the Belize collection and our collection, females of *Psychodopygus panamensis* had blood-fed. Finally, we found *Bichromomyia flaviscutellata* (Mangabeira Filho 1942) in the Los Guácharos cave, agreeing with a previous report in Brazilian caves that reported this species can have a cavernicolous habitat (Alves et al. 2011).

#### 3.4. Species richness and comparisons to other caves

Our sampling yielded 52 individuals and revealed the existence of 17 species in the Los Guácharos cave. This sample has one of the highest species richness in any Neotropical sample collected in caves (Table 2). We also calculated the Shannon diversity index for each location and a normalized evenness index for 32 caves sampled in the Neotropics. The Guacharos cave also has the highest Shannon's Index (*H*) and the one of the highest evenness indexes of any Neotropical cave (Table 2) suggesting one of the highest diversity of sandflies in Neotropical caves. Nonetheless, our sample has a relatively low sample size compared to others. To account for differences in the sample size, we rarefied two of the diversity indexes and the results are qualitatively similar to those before rarefaction (Table 2). Regardless of the species metric diversity, the Los Guácharos cave has a high species diversity compared to other samples.

Next, we tested whether the diversity of sandflies in caves followed a latitudinal gradient of diversity by regressing the latitude of the collection with each of the three metrics of diversity (Fig. 3). Contrary to our expectation, we find that none of the regressions is significant suggesting that species diversity of sandfly caves does not decrease as the cave gets farther from the Equator. The result was the same with any of the three diversity indexes regardless of whether we corrected for effort

**Table 1**  
List of Phlebotomines collected in the Los Guácharos Cave in the Rio Claro Natural reserve (Antioquia), Colombia. F= female; M= male. \* = species collected in Colombia for the first time. The number within the parentheses shows the number of gravid females or males with rotated genitalia (reproductive stage).

Species	Cave		Rock-breaks		Fold		Vegetation		Total
	F	M	F	M	F	M	F	M	
<i>Pintomyia ovallesi</i>	0	0	6 (1)	0	3(2)	0	0	0	9 (17.3%)
<i>Lutzomyia gomezi</i>	0	0	1(1)	0	3(3)	0	2(2)	0	6 (11.53%)
<i>Warileya hertigi</i>	5	5	1 (1)	0	0	0	0	0	11 (21.14%)
<i>Micropygomyia trinidadensis</i>	0	0	2(1)	0	0	0	1	1	4 (7.69%)
<i>Psathyromyia carpenteri</i>	0	0	3 (3)	1	0	0	0	0	4 (7.69%)
<i>Psychodopygus panamensis</i>	0	1	0	0	1	1	0	0	3 (5.7%)
<i>Brumptomyia hamata</i>	0	1	0	1	0	0	0	0	2 (3.84%)
<i>Evandromyia saulensis</i>	0	0	0	0	1	0	1	0	2 (3.84%)
<i>Helcorcitomyia sp.</i>	1 (1)	0	1	0	0	0	0	0	2 (3.84%)
<i>Lutzomyia hartmanni</i>	1 (1)	0	1	0	0	0	0	0	2 (3.84%)
<i>Bichromomyia flaviscutellata</i>	0	0	0	0	1 (1)	0	0	0	1 (1.92%)
<i>Bichromomyia olmeca nociva*</i>	0	0	0	0	0	1	0	0	1 (1.92%)
<i>Lutzomyia sanguinaria</i>	0	0	0	0	1	0	0	0	1 (1.92%)
<i>Micropygomyia atroclavata</i>	0	0	0	1	0	0	0	0	1 (1.92%)
<i>Micropygomyia micropyga</i>	0	0	0	0	0	1	0	0	1 (1.92%)
<i>Pressatia compositi</i>	0	0	0	0	0	1	0	0	1 (1.92%)
<i>Warileya sp.</i>	0	0	0	1	0	0	0	0	1 (1.92%)
Total Juveniles(reproductive)	7(2)	7	15 (7)	4	10(6)	4	4 (2)	1	52 (100%)
<b>Total (%)</b>	<b>14 (26.9%)</b>		<b>19 (36.53%)</b>		<b>14 (26.90%)</b>		<b>5 (9.61%)</b>		

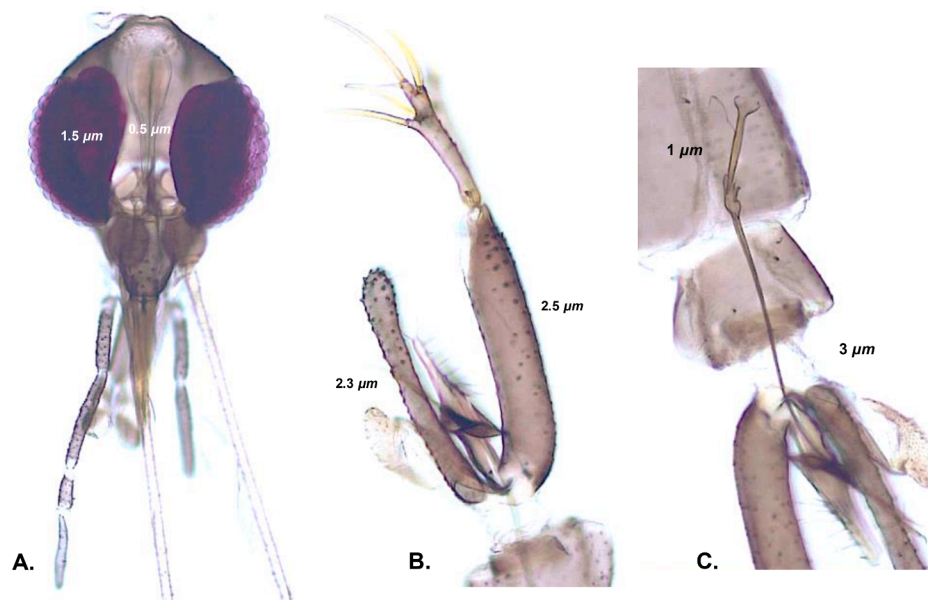


Fig. 2. Diagnostic traits of the new Phlebotomine register for the Colombian territory. *Bichromomyia olmeca nociva*: (a) Head and interocular space, (b) Epandrial lobes slightly shorter than gonocoxite, (c) spermiducts/sperm pump.

Table 2

Compilation of neotropical studies of sandfly cave diversity and their respective metrics of species diversity. H: Shannon’s H index. J: Evenness. \*Includes estimates over multiple years. & includes more than one connected cave.

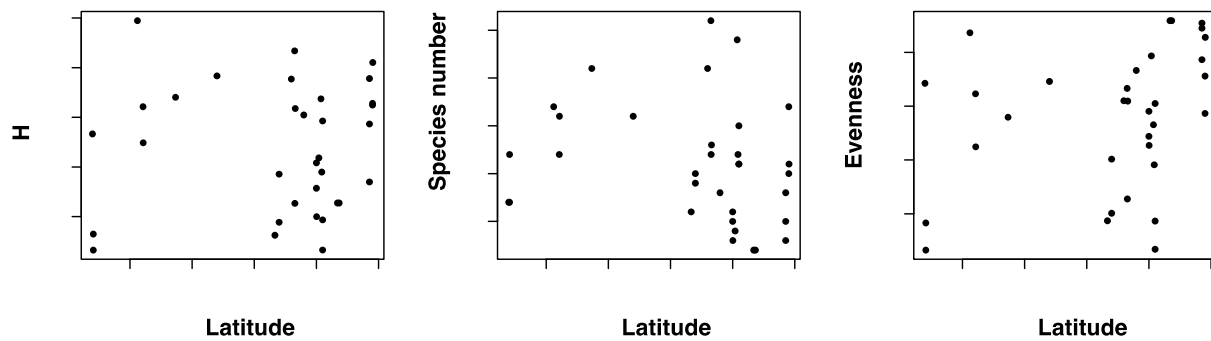
Locality	Reference	Before rarefaction			After rarefaction(1,000 replicates; 6 species)	
		H	Species richness	Evenness	H	Evenness
Los Guácharos Cave, Colombia	This study	2.474	17	0.873	1.560	0.448
Monte Cristo, Brazil	(Barata et al., 2012)	1.589	13	0.619	0.693	0.253
Salitre, Brazil	(Barata et al., 2012)	0.633	12	0.255	1.011	0.506
North Rondônia, Brazil	(Ogawa et al. 2016)	1.702	21	0.559	1.011	0.443
South Rondônia, Brazil	(Ogawa et al. 2016)	1.918	16	0.692	1.330	0.253
Gruta dos Animais, Brazil	(Alves et al., 2011)	0.163	12	0.065	0.000	0.434
Gruta do Maruaga, Brazil	(Alves et al., 2011)	0.324	7	0.166	0.000	0.225
Gruta dos Lages, Brazil	(Alves et al., 2011)	1.333	7	0.685	0.451	0.366
Millionario, Belize *	(Williams 1976)	0.312	6	0.174	0.451	0.000
Augustine, Belize	(Williams 1976)	0.443	9	0.201	0.637	0.000
SA, Belize	(Williams 1976)	0.928	10	0.403	0.451	0.347
Bodoquera, Brazil	(Galati et al., 2003)	0.949	12	0.382	1.011	0.434
Pitangueiras, Brazil	(Galati et al., 2003)	0.163	11	0.068	0.000	0.225
Anhumas, Brazil	(Galati et al., 2003)	0.467	15	0.173	0.000	0.225
Guaicurus, Brazil	(Galati et al., 2003)	1.463	11	0.610	1.330	0.896
Jardim, Brazil	(Galati et al., 2003)	0.637	2	0.918	0.636	0.318
Yucatan, Mexico	(Oca-Aguilar et al., 2013)	1.041	6	0.581	0.868	0.506
Ibitipoca, Brazil	(Carvalho et al., 2011)	0.637	2	0.918	0.451	0.347
Corumba, Brazil	(Galati et al., 1997)	1.524	8	0.733	1.561	0.253
Colorida, Brazil	(Galati et al., 2010b)	1.891	8	0.909	1.011	0.311
Minotauro, Brazil	(Galati et al., 2010b)	1.433	5	0.890	1.099	0.332
Barra bonita, Brazil	(Galati et al., 2010)	0.849	3	0.773	0.637	0.212
Santana, Brazil	(Galati et al., 2010b)	2.053	11	0.856	1.561	0.780
Couto, Brazil	(Galati et al., 2010b)	1.640	10	0.712	1.242	0.414
Morro Preto, Brazil	(Galati et al., 2010b)	1.623	17	0.573	0.451	0.665
Moeda Sul, Brazil	(Campos et al., 2020)	1.091	4	0.787	1.011	0.337
Rola Moça 39, Brazil	(Campos et al., 2020)	0.786	5	0.488	0.451	0.451
Rola Moça 40, Brazil	(Campos et al., 2020)	0.499	3	0.454	0.451	0.000
Lassance, Brazil	(Carvalho et al., 2013)	1.886	21	0.620	1.561	0.443
Pitoco, Brazil	(de Almeida et al., 2019)	2.169	26	0.666	1.561	0.443
Parauapebas, Brazil	(Teodoro et al. 2021)	1.243	16	0.449	1.561	0.665
Curionópolis, Brazil	(Teodoro et al. 2021)	1.606	12	0.646	1.099	0.253
Pains, Brazil &	(Campos et al., 2017)	1.686	24	0.531	1.330	0.113

or not (Table 3).

### 3.5. Phylogenetic signal of cavernicolous habitat

We used the Psychodidae phylogenetic tree to study whether the cavern habitat association had a phylogenetic signal in the family. Our

metric of phylogenetic signal was close to zero (Pagel’s  $\lambda = 6.611 \times 10^{-5}$ ;  $\log Lik = -45.273$ ) which suggests no effect of the phylogeny on the distribution of cavernicolous habitat. The distribution of the trait along the tree suggests the same pattern as cavernicolous species are not related to each other (Fig. 4). Thus cave use likely evolved at least three times in the family as it appears in at least three different genera.



**Fig. 3.** Three different metrics of the species diversity in Neotropical sandfly cave collections. Left. Shannon's  $H$  index Center. Number of species Right. Evenness. Each point represents the species richness metric for a sampling listed in Table 2. Significance of the regressions is shown in Table 3. None of the three metrics shows evidence for a latitudinal gradient of sandfly species diversity collected in caves.

**Table 3**

Linear models show no evidence of a latitudinal gradient of species diversity for cave sandflies.

Diversity index	Estimate	Standard Error	t-value	P-value
$H$	$3.099 \times 10^{-3}$	0.016	0.193	0.848
Species number	-0.185	0.155	-1.196	0.241
Evenness	0.011	$6.215 \times 10^{-3}$	1.774	0.086
$H$ (rarefied)	$6.672 \times 10^{-3}$	0.013	0.500	0.620
Evenness (rarefied)	$9.322 \times 10^{-4}$	$5.13 \times 10^{-3}$	0.182	0.857

#### 4. Discussion

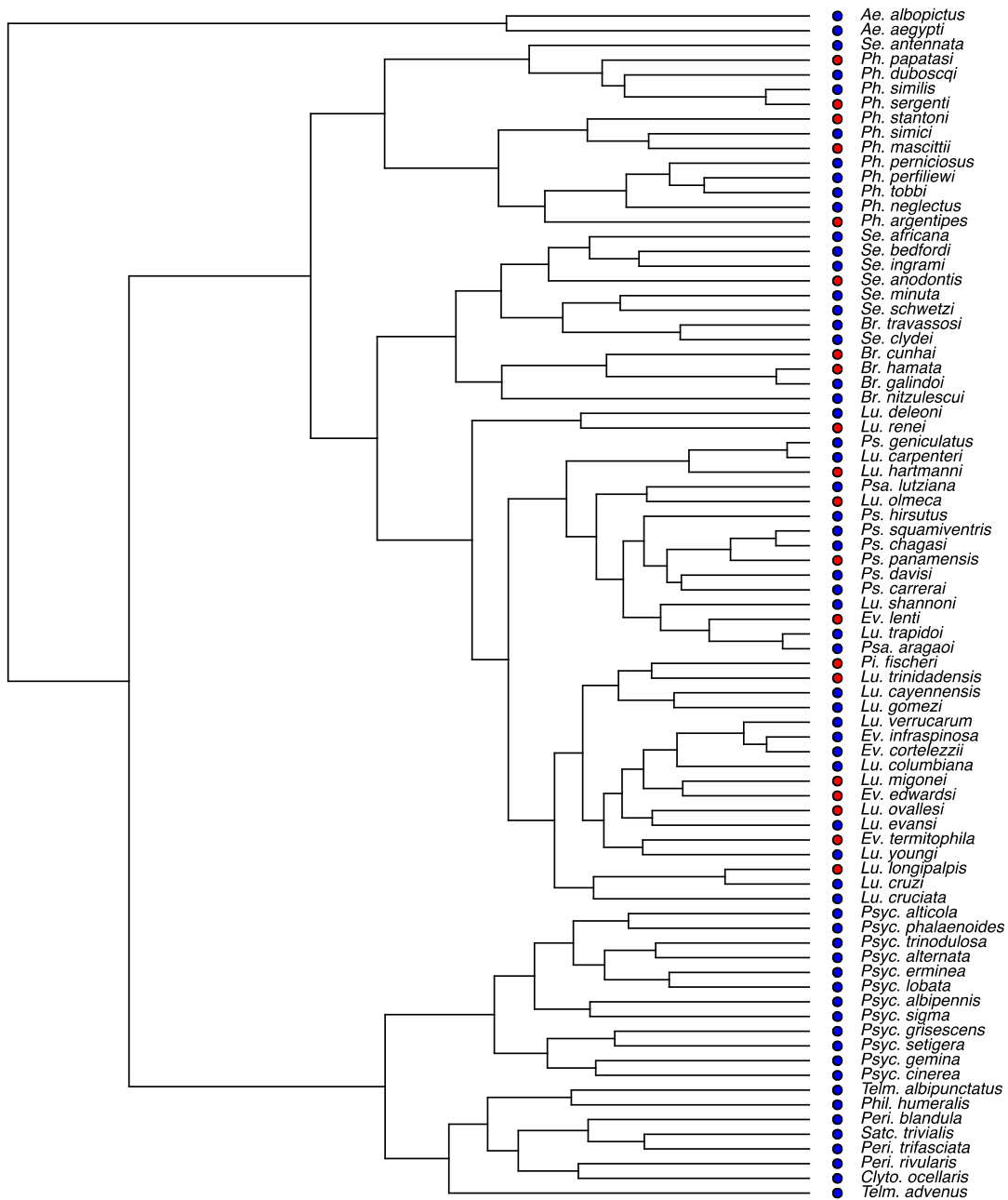
The study of the habitat of vector species can yield important information on potential control strategies to limit their ability to transmit disease. We did a point sampling of a cave in western Colombia. Our report is the first multispecies sampling of a cave in the northern Andes. We report that a *Bichromomyia olmeca* has a larger geographic range than previously reported (Shimabukuro et al., 2017; de Melo et al. 2020). The species is naturally infected and can effectively transmit *Leishmania amazonensis* (Brazil et al. 2015), which highlights the importance of finding this species in cave systems relatively close to human settlements and with frequent ecotourism. *Bichromomyia olmeca* is found throughout Central America to Brazil (GBIF: DOI: <https://doi.org/10.15468/dl.g77tbs>, years: 1953–2018, Table S1 in Moo-Llanes et al., 2013, and Galati 2018) and seems to be a species complex composed of at least four different lineages: *B. o. nociva* (reported in this study), and *B. o. olmeca* (mainly localized in Mexico), *B. o. bicolor* (reported in Panamá, Colombia, and Brazil), and *B. flaviscutellata sensu stricto* (de Melo et al. 2020). The Guacharos cave harbors both *B. olmeca nociva* and *B. flaviscutellata* and adds to the list of locales where lineages of the *B. olmeca* species complex coexist in the same geographic range (Fairchild and Theodor 1971; Escobar Vasco 1989; Romero Ricardo et al. 2013; de Melo et al. 2020; Santatana et al. 2020). The identification of *B. olmeca nociva* in Colombia poses the need to revisit the geographic range of each species, with a special emphasis on areas of coexistence which might facilitate gene flow between vector species.

Determining range limits has practical implications for our understanding of vector biology. Under current global warming scenarios, *B. olmeca* is predicted to expand its current range to subtropical and temperate areas (Moo-Llanes et al., 2013). This prediction was only possible after a systematic habitat characterization of the species habitat (108 collection sites, Table 1 in (Moo-Llanes et al., 2013) which in turn revealed the suitable current and future ecological niche of the species. To date, the habitat characterization of *B. olmeca nociva* or other species in the species complex, —at least to the same detail as *B. o. olmeca*, —is lacking. We hypothesize that lack of systematic sampling in the Northern Andes might explain the discontinuous geographic range of other species.

More generally, we used the data from our sampling to infer ecological and evolutionary patterns about cave habitat use in sandflies. We find that the Los Guácharos cave has the largest species diversity of any sampled Neotropical cave. Opposite to our expectations, this analysis revealed that sandfly cave species diversity does not follow a latitudinal gradient in the South American continent. Latitudinal gradients, in which the tropics are the most diverse locales, have been reported for almost all systematically sampled taxa (reviewed in Mannion et al., 2014; Jablonski et al., 2017; exceptions listed in Kindmann et al., 2007). Nonetheless, the drivers of the pattern are hotly debated and are likely to be caused by a variety of processes. Fish for example show higher diversity in the tropics (Mittelbach et al., 2007; Stuart-Smith et al., 2013) but a higher speciation rate in temperate areas (Rabosky et al., 2018). Our results find that the variance of species richness might be the largest in the tropics, as the samples close to the equator show one of the highest (Los Guácharos cave, Colombia; this study) and some of the lowest species richness (Gruta dos Animais, Amazonas, Brazil; Alves et al., 2011) of all the sampled caves. More systematic sampling will be needed before the hypothesis that the tropics have a larger variance in species richness across localities, but the possibility is tantalizing.

Finally, we assessed the evolution of cave habitat choice across the family Psychodidae. We find no phylogenetic signal for cave-habitat association, which supports the possibility that habitat association has evolved repeatedly in the family. Other metrics of the ecological niche of sandflies are highly conserved in the sandfly phylogeny. Proxies of climatic niche are more conserved than expected by the phylogeny (D'Agostino et al., 2022), suggesting that niche conservatism exists within the family for certain traits. A different possibility is that many species are generalists that can use caves as a habitat when present, but limited sampling makes verifying this a challenging hypothesis to test. In this scenario, species are only cavernicolous when there are available caves nearby. The current sampling does not allow us to differentiate between these possibilities. Mark recapture experiments releasing sandflies at the entrance of caves and determining their habitat choice should reveal whether the collection of species in caves is due to happenstance. Regardless of the level of niche conservation of cave use in the family, caves seem to be a niche that is used widely in sandflies and their ecological characterization might be of importance for understanding disease dynamics and vector control.

Our study has caveats that are worth pointing out. First, our sampling is not systematic and did not address the variation of diversity along the year. A proper systematic sampling that addresses species diversity seasonally (at least in the dry and wet seasons) will be necessary to understand the true potential of Colombian caves as reservoirs of vector species. The limitation of systematic sampling extends to other studies (but not all; e.g., Carvalho et al., 2013), and has implications to our inferences of the patterns and processes that lead to the phylogenetic signal of cave species. We note that our results remain qualitatively the same after accounting for this lack of systematic sampling by correcting



**Fig. 4. Phylogenetic signal of cavernicolous habitat in Psychodidae.** The dots at the tree tips show the extant known habitat choice for each species in the phylogeny. Ae: *Aedes*; Se: *Sergentomyia*; pH: *Phlebotomus*; Br: *Brumptomyia*; Lu: *Lutzomyia*; Ps: *Psychodopygus*; Psa: *Psathromyia*; Ev: *Evandromyia*; Pi: *Pintomyia*; Psc: *Psychoda*; Telm: *Telmatoscopus*; Phil: *Philosepedon*; Satc: *Satchelliella*; Peri: *Pericoma*; Clyto: *Clytocerus*. Blue: not known to be associated with caves, Red: associated with caves.

for sampling effort. Despite this, a more expanded sampling of different caves in the Northern Andes is warranted. One additional caveat is that we expect the phylogeny of sandflies to change as data for more species and better genetic markers become available. The current topology is based on four loci and only includes 79 Psychodidae species. In all certainty, years to come will see an increase on the amount of genomic data in sandflies and this will lead to a revision of the phylogenetic relationships between these species, including some that have cavernicolous habitats. Our estimates of phylogenetic signal will thus change slightly. However, our conclusion that cavernicolous habitat has evolved in several instances in the phylogeny is likely to stand unless the

topology is dramatically revised. Our results confirm the need for thorough and systematic sampling of sandfly diversity, including habitats that have not often been sampled (e.g., caves), in order to better understand this ecologically and medically important clade.

**CRedit authorship contribution statement**

**Manuela Velásquez Londoño:** Investigation, Formal analysis. **Adam M.M. Stuckert:** Investigation, Visualization, Writing – review & editing, Formal analysis. **Rafael J. Vivero:** Conceptualization, Investigation, Formal analysis. **Daniel R. Matute:** Conceptualization,

Investigation, Visualization, Writing – original draft, Formal analysis, Funding acquisition.

## Declaration of Competing Interest

The authors declare no competing interests.

## Acknowledgements

We thank the Matute lab for constructive feedback on the manuscript. This work was supported by the National Science Foundation (Dimensions of Biodiversity award 1737752 to D.R.M.). The funders had no role in any aspect of study design, data collection and analysis, or decisions with respect to publication.

## References

- Acevedo, M., de los, Á., Arrivillaga, J., 2008. Eco-epidemiología de flebovirus (Bunyaviridae, Phlebovirus) transmitidos por flebotomos (Psychodidae, Phlebotominae). *Boletín de Malariología y Salud ambiental* 48, 3–16.
- Alexander, B., 2000. Sampling methods for phlebotomine sandflies. *Med. Vet. Entomol.* 14, 109–122. <https://doi.org/10.1046/j.1365-2915.2000.00237.x>.
- Alvar, J., Vélez, I.D., Bern, C., Herrero, M., Desjeux, P., et al., 2012. Leishmaniasis worldwide and global estimates of its incidence. *PLoS One* 7, e35671.
- Alves, V.R., R.A.de Freitas, Barrett, T., 2008. *Lutzomyia maruaga* (Diptera: psychodidae), a new bat-cave sand fly from Amazonas. Brazil. *Memórias do Instituto Oswaldo Cruz* 103, 251–253.
- Alves, V.R., de Freitas, R.A., Santos, F.L., Barrett, T.V., 2011. Diversity of sandflies (Psychodidae: phlebotominae) captured in sandstone caves from Central Amazonia. Brazil. *Memórias do Instituto Oswaldo Cruz* 106, 353–359.
- Almeida, P.S., Paula, M.B., Brilhante, A.F., Medeiros-Sousa, A.R., Neitzke-Abreu, H.C., Carriho, C.J.S., Costa, P.C., Galati, E.A.B., 2019. Phlebotomine (Diptera: psychodidae) fauna in a cavern containing cave paintings and its surrounding environment, Central-West Brazil. *Acta Trop.* 199, 105–151, 2019.
- Bailey, F., Mondragon-Shem, K., Hotez, P., Ruiz-Postigo, J.A., Al-Salem, W., et al., 2017. A new perspective on cutaneous leishmaniasis—Implications for global prevalence and burden of disease estimates. *PLoS Negl. Trop. Dis.* 11, e0005739 <https://doi.org/10.1371/journal.pntd.0005739>.
- Barata, R.A., Serra e Meira, P.C.L., Carvalho, G.M.de L., 2012. *Lutzomyia diamantinensis* sp. nov., a new phlebotomine species (Diptera: psychodidae) from a quartzite cave in Diamantina, state of Minas Gerais. Brazil. *Memórias do Instituto Oswaldo Cruz* 107, 1006–1010.
- Batieha, A., Saliba, E.K., Graham, R., Mohareb, E., Hijazi, Y., et al., 2000. Seroprevalence of West Nile, Rift Valley, and sandfly arboviruses in Hashimiah. *Jordan. Emerg. Infectious Dis.* 6, 358.
- Bejarano, E.E., Estrada, L.G., 2016. Family psychodidae. *Zootaxa* 4122, 187–238.
- Bejarano, E.E., Vivero, R.J., Uribe, S., 2018. Primer registro del flebotomíneo *Warileya (Hertigia) hertigi* en Antioquia. *Colombia. Biomédica* 38, 277–281.
- Blavier, A., Laroche, L., Randrianambinintsoa, F.J., Lucas, V., Gantier, J.-C., et al., 2019. Phlebotomine sandflies (Diptera, Psychodidae) from the Ankarana tsingy of northern Madagascar: inventory and description of new taxa. *Parasite* 26.
- Brazil, R.P., Rodrigues, A.A.F. Andrade-Filho, J.D. (2015) Sand fly vectors of *Leishmania* in the Americas A mini review. *Entomology, Ornithology and Herpetology: Current Research*, 04, 4–7. <https://doi.org/10.4172/2161-0983.1000144>.
- Caceres, A.G., Galati, E.A., Le Pont, F., Velásquez, C., 1997. Possible role of *Lutzomyia maranonensis* and *Lutzomyia robusta* (Diptera: psychodidae) as vectors of human bartonellosis in three provinces of region nor Oriental del Marañón. *Peru. Rev. Inst. Med. Trop. Sao Paulo* 39, 51–52. <https://doi.org/10.1590/s0036-46651997000100011>.
- Campos, A.M., Dos Santos, C.L.C., Stumpp, R., Da Silva, L.H.D., Maia, R.A., et al., 2017. Photoperiod Differences in Sand Fly (Diptera: psychodidae) Species Richness and Abundance in Caves in Minas Gerais State. *Brazil. J. Med. Entomol.* 54, 100–105. <https://doi.org/10.1093/jme/tjw135>.
- Campos, A.M., Maia, R.A., Capucci, D., Paglia, A.P., Andrade, J.D., 2020. Species composition of sand flies (Diptera: psychodidae) in caves of Quadrilátero Ferrífero, state of Minas Gerais, Brazil. *PLoS One* 15 (3), e0220268, 2020.
- Carvalho, G.M., De Vasconcelos, F.B., Da Silva, D.G., Botelho, H.A., Filho, J.D.A., 2011a. Diversity of phlebotomine sand flies (Diptera: psychodidae) in Ibitipoca State Park, Minas Gerais, Brazil. *J. Med. Entomol.* 48, 764–769.
- Carvalho, G.M.de L., Brazil, R.P., Ramos, M.C.das N.F., Serra e Meira, P.C.L., Zenóbio, A. P.L.de A., et al., 2013. Ecological aspects of phlebotomine sandflies (Diptera: psychodidae) from a cave of the speleological province of Bambuí. *Brazil. PLoS One* 8, e71158.
- Carvalho, G.M.L., Brazil, R.P., Rêgo, F.D., Ramos, M.C.N.F., Zenóbio, A.P.L.A., et al., 2017. Molecular Detection of Leishmania DNA in Wild-Caught Phlebotomine Sand Flies (Diptera: psychodidae) From a Cave in the State of Minas Gerais. *Brazil. J. Med. Entomol.* 54, 196–203. <https://doi.org/10.1093/jme/tjw137>.
- Comer, J.A., Tesh, R.B., 1991. Phlebotomine sand flies as vectors of vesiculoviruses: a review. *Parassitologia* 33, 143–150.
- Cooper, N., Jetz, W., Freckleton, R.P., 2010. Phylogenetic comparative approaches for studying niche conservatism. *J. Evol. Biol.* 23, 2529–2539 <https://doi.org/10.1111/j.1420-9101.2010.02144.x>.
- Cordeiro D.P., and R. Wagner, 2018 Chapter 16.10 - Family Psychodidae, pp. 765–770 in *Thorp and Covich's Freshwater Invertebrates (Fourth Edition)*, edited by Hamada N., Thorp J. H., Rogers D. C. Academic Press.
- Costa, J.C.R., Marchi, G.H., Santos, C.S., Andrade, M.C.M., Junior, S.C., et al., 2021. First molecular evidence of frogs as a food source for sand flies (Diptera: phlebotominae) in Brazilian caves. *Parasitol. Res.* 120, 1571–1582.
- Cutolo, A.A., Camargo, D.A., Cutolo, A.A., Von, C.J., Galati, E.A.B., 2008. *Lutzomyia longipalpis* (Diptera, Psychodidae) em Cuesta Basáltica, na bacia hidrográfica do Rio Corumbataí, Região Centro-leste do Estado de São Paulo. *Rev. Bras. Epidemiol.* 11 (2), 336–339.
- D'Agostino E.R.R., Vivero, R., Romero, L., Bejarano, E., Hurlbert A.H., Comeault A.A., Matute, D.R. 2022. Climatic Niche Conservatism in a Clade of Disease Vectors (Diptera: Phlebotominae). *bioRxiv.* 475052.
- de Melo, L.B., Alencar, R.B., Scarpassa, V.M., 2020. Molecular taxonomy and phylogenetic inferences of *Bichromomyia flaviscutellata* complex based on the COI gene DNA barcode region. *Infect. Genet. Evol.* 81, 104256.
- Depaquit, J., Grandadam, M., Fouque, F., Andry, P.E., Peyrefitte, C., 2010. Arthropod-borne viruses transmitted by Phlebotomine sandflies in Europe: a review. *Eurosurveillance* 15, 19507.
- Dionisio, D., Esperti, F., Vivarelli, A., Valassina, M., 2003. Epidemiological, clinical and laboratory aspects of sandfly fever. *Curr. Opin. Infect. Dis.* 16, 383–388.
- Escobar Vasco J.P., 1989 Variación estacional, distribución vertical y actividad rodentofílica de *Lutzomyia olmeca bicolor* y *Lu. panamensis* (Diptera: psychodidae) en Gamboa-Panamá.
- Espindola, A., Buerki, S., Jacquier, A., Ježek, J., Alvarez, N., 2012. Phylogenetic relationships in the subfamily Psychodinae (Diptera, Psychodidae). *Zool. Scr.* 41, 489–498 <https://doi.org/10.1111/j.1463-6409.2012.00544.x>.
- Fairchild, G.B., Hertig, M., 1957. Notes on the *Phlebotomus* of Panama XIII. The *Vexator* group, with descriptions of new species from Panama and California. *Ann. Entomol. Soc. Am.* 50, 325–334.
- Fairchild, G.B., Theodor, O., 1971. On *Lutzomyia flaviscutellata* (Mangabeira) and *L. olmeca* (Vargas and Diaz-Najera) (Diptera: psychodidae). *J. Med. Entomol.* 8, 153–159.
- Feliciangeli, M.D., 1997. Hourly activity of *Lutzomyia ovallesi* and *L. gomezi* (Diptera: psychodidae), vectors of cutaneous leishmaniasis in northcentral Venezuela. *J. Med. Entomol.* 34, 110–115.
- FELICIANGELI, D.M., Rabinovich, J., 1998. Abundance of *Lutzomyia ovallesi* but not *Lu. gomezi* (Diptera: psychodidae) correlated with cutaneous leishmaniasis incidence in north-central Venezuela. *Med. Vet. Entomol.* 12, 121–131.
- Ferro, C., López, M., Fuya, P., Lugo, L., Cordovez, J.M., et al., 2015. Spatial Distribution of Sand Fly Vectors and Eco-Epidemiology of Cutaneous Leishmaniasis Transmission in Colombia. *PLoS One* 10, e0139391. <https://doi.org/10.1371/journal.pone.0139391>.
- Fox, J., Weisberg, S., Adler, D., Bates, D., Baud-Bovy, G., Ellison, S., Firth, D., Friendly, M., Gorjanc, G., Graves, S. and Heiberger, R., 2012. Package 'car'. Vienna: R Foundation for Statistical Computing, 16.
- Fox, J., Weisberg, S., 2019. Companion to Applied Regression, Third Edition. Thousand Oaks: Sage.
- Galati, E.A., 2018. Phlebotominae (Diptera, Psychodidae): classification, morphology and terminology of adults and identification of American taxa. *Brazilian Sand Flies.* Springer, Cham, pp. 9–212.
- Galati, E.A., Nunes, V.L., Boggiani, P.C., Dorval, M.E.C., Cristaldo, G., et al., 2003. Phlebotomines (Diptera, Psychodidae) in caves of the Serra da Bodoquena, Mato Grosso do Sul State, Brazil. *Rev. Bras. Entomol.* 47, 283–296.
- Galati, E.A.B., 2019. Morfologia e Terminologia De Phlebotominae (Diptera: Psychodidae). *Classificação e identificação De Taxons Das Americas.* Apostila da Disciplina Bioecologia e Identificação de Phlebotominae. Programa de Pós-Graduação em Saúde Pública. São Paulo: Faculdade de Saúde Pública da Universidade de São Paulo.
- Galati, E.A.B., Nunes, V.L., Rego, F.A., Oshiro, E.T., Chanf, M.R., 1997. Estudo de flebotomíneos (Diptera: psychodidae) em foco de leishmaniose visceral no Estado de Mato Grosso do Sul. *Brasil. Rev. Saúde Pública* 31 (4), 378–390.
- Galati, E.A.B., Marrasa, A.M., Andrade, R.M., Consales, C.A., Bueno, E.M., 2010. Phlebotomines (Diptera, Psychodidae) in the Speleological Province of the Ribeira Valley - 1. Parque Estadual Intervaless, state of São Paulo. *Brazil. Rev. Bras. Entomol.* 54 (2), 311–321.
- González, R., Sousa, L.D., Devera, R., Jorquera, A., Ledezma, E., 1999. Seasonal and nocturnal domiciliary human landing/biting behaviour of *Lutzomyia (Lutzomyia) evansi* and *Lutzomyia (Psychodopygus) panamensis* (Diptera: Psychodidae) in a periurban area of a city on the Caribbean coast of eastern Venezuela (Barcelona; Anzoátegui State). *Trans. R. Soc. Trop. Med. Hyg.* 93, 361–364. [https://doi.org/10.1016/S0035-9203\(99\)90115-3](https://doi.org/10.1016/S0035-9203(99)90115-3).
- Harmon, L.J., Weir, J.T., Brock, C.D., Glor, R.E., Challenger, W., 2008. GEIGER: investigating evolutionary radiations. *Bioinformatics* 24, 129–131.
- Ibáñez-bernal, S., 1999. Phlebotominae (Diptera: psychodidae) de Mexico. 1.- *Brumptomomyia frança* y Parrot; *Lutzomyia frança*, las especies de *L. (Lutzomyia) frança* y del grupo *verrucarum*. *Folia Entomol. Mex* 107, 116.
- Izri, A., Temmam, S., Moureau, G., Hamrioui, B., De Lamballerie, X., et al., 2008. Sandfly fever Sicilian virus. *Algeria. Emerg. Infectious Dis.* 14, 795.
- Jablonski, D., Huang, S., Roy, K., Valentine, J.W., 2017. Shaping the Latitudinal Diversity Gradient: new Perspectives from a Synthesis of Paleobiology and Biogeography. *Am. Nat.* 189, 1–12. <https://doi.org/10.1086/689739>.



- Karimkhani, C., Wanga, V., Coffeng, L.E., Naghavi, P., Dellavalle, R.P., et al., 2016. Global burden of cutaneous leishmaniasis: a cross-sectional analysis from the Global Burden of Disease Study 2013. *Lancet Infect. Dis.* 16, 584–591 [https://doi.org/10.1016/S1473-3099\(16\)00003-7](https://doi.org/10.1016/S1473-3099(16)00003-7).
- Killick-Kendrick, R., 1990. Phlebotomine vectors of the leishmaniasis: a review. *Med. Vet. Entomol.* 4, 1–24.
- Kindlmann, P., Schödelbauerová, I., Dixon, A.F., 2007. Inverse Latitudinal Gradients in Species diversity. *Scaling Biodiversity*. Cambridge University Press, Cambridge, UK.
- Mangabeira Filho, O., 1942. 13ª Contribuição ao estudo dos *Flebotomus* (Diptera: psychodidae): *flebotomus (Brumptomyia) travassosi* Mangabeira, 1942. *Memórias do Instituto Oswaldo Cruz* 37, 375–381.
- Mannion, P.D., Upchurch, P., Benson, R.B., Goswami, A., 2014. The latitudinal biodiversity gradient through deep time. *Trends Ecol. Evol. (Amst.)* 29, 42–50.
- Memmott, J., 1991. Sandfly distribution and abundance in a tropical rain forest. *Med. Vet. Entomol.* 5, 403–411.
- Memmott, J., 1992. Patterns of sandfly distribution in tropical forest: a causal hypothesis. *Med. Vet. Entomol.* 6, 188–194.
- Mittelbach, G.G., Schemske, D.W., Cornell, H.V., Allen, A.P., Brown, J.M., et al., 2007. Evolution and the latitudinal diversity gradient: speciation, extinction and biogeography. *Ecol. Lett.* 10, 315–331.
- Moncada, L.E., Pineda, F., Muñoz, J., Ferreira, G., 1989. Estudio de la presencia de *Histoplasma capsulatum* en la tierra de cuatro cuevas localizadas en la región del Río Claro (Antioquia). *Iatreia* 2 (3), 195–200.
- Moo-Llanes, D., Ibarra-Cerdeña, C.N., Rebollar-Téllez, E.A., Ibáñez-Bernal, S., González, C., et al., 2013. Current and future niche of North and Central American sand flies (Diptera: psychodidae) in climate change scenarios. *PLoS Negl. Trop. Dis.* 7, e2421.
- Munstermann L.E., 2019 Chapter 12 - Phlebotomine Sand Flies and Moth Flies (Psychodidae), pp. 191–211 in *Medical and Veterinary Entomology (Third Edition)*, edited by Mullen G. R., Durden L. A. Academic Press.
- Nitzulescu, V., 1931. Essai de classification des phlébotomes. *Annales de Parasitologie humaine et comparée* 9, 271–275.
- Oca-Aguilar, A.C.M.de, Moo-Llanes, D., Rebollar-Téllez, E.A., 2013. Sand Fly Species from a Karstic Cave in the Peninsula of Yucatan, Mexico. *entn* 123, 191–200. <https://doi.org/10.3157/021.123.0305>.
- Ogawa, G.M., Pereira Júnior, A.M., Resadore, F., Ferreira, R.D.G.M., Medeiros, J.F., Camargo, L.M.A., 2016. Sandfly fauna (Diptera: Psychodidae) from caves in the state of Rondônia, Brazil. *Revista Brasileira de Parasitologia Veterinária*, 25, pp. 61–68.
- Oksanen, J., 2013. *Vegan: ecological diversity*, p. 368.
- Oksanen, J., Blanchet, F.G., Kindt, R., Legendre, P., Minchin, P.R., et al., 2013. Package 'vegan'. *Bull. Trimest. Plan. Fam.* 1–295.
- Oksanen, J., Blanchet, F.G., Kindt, R., Legendre, P., Minchin, P.R., et al., 2015. *Vegan Community Ecology package: Ordination methods, Diversity Analysis and Other Functions For Community and Vegetation Ecologists*. R package ver 2–3.
- Okwor, I., Uzonna, J., 2016. Social and economic burden of human leishmaniasis. *Am. J. Trop. Med. Hyg.* 94, 489.
- Ortiz, I., 1951. descripción de dos nuevas especies del genero *Phlebotomus rondani*. *Acta biológica venezolica* 231.
- Pagel, M., 1997. Inferring evolutionary processes from phylogenies. *Zool. Scr.* 26, 331–348.
- Pagel, M., 1999. Inferring the historical patterns of biological evolution. *Nature* 401, 877–884.
- Paradis, E., Claude, J., Strimmer, K., 2004. APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* 20, 289–290. <https://doi.org/10.1093/bioinformatics/btg412>.
- Paradis, E., Schliep, K., 2019. ape 5.0: an environment for modern phylogenetics and evolutionary analyses in R. *Bioinformatics* 35, 526–528.
- Pave, H., Blagoderov, V., Mostovski, M., 2011. Animal biodiversity: an outline of higher-level classification and survey of taxonomic richness. *Order Diptera Linnaeus. Zootaxa* 3148, 222–229.
- Pennell, M.W., Eastman, J.M., Slater, G.J., Brown, J.W., Uyeda, J.C., et al., 2014. geiger v2. 0: an expanded suite of methods for fitting macroevolutionary models to phylogenetic trees. *Bioinformatics* 30, 2216–2218.
- Phumea, A., Wacharapluesadee, S., Petcharat, S., Tawatsin, A., Thavara, U., et al., 2021. Detection of Changuinola virus (Reoviridae: orbivirus) in field-caught sand flies in southern Thailand. *Trans. R. Soc. Trop. Med. Hyg.*
- Pontarp, M., Bunnefeld, L., Cabral, J.S., Etienne, R.S., Fritz, S.A., et al., 2019. The latitudinal diversity gradient: novel understanding through mechanistic eco-evolutionary models. *Trends Ecol. Evol. (Amst.)* 34, 211–223.
- Quate, L.W., 1962. The Psychodidae of Batu Caves. *Malaya (Diptera). Pacific Insects* 4, 219–234.
- R Core Team, 2021 R: A language and Environment For Statistical Computing. R Foundation for Statistical Computing, Vienna, Austria. URL <https://www.R-project.org/>.
- Rabinovich, J.E., Feliciangeli, M.D., 2004. Parameters of *Leishmania braziliensis* transmission by indoor *Lutzomyia ovallesi* in Venezuela. *Am. J. Trop. Med. Hyg.* 70, 373–382.
- Rabosky, D.L., Chang, J., Title, P.O., Cowman, P.F., Sallan, L., et al., 2018. An inverse latitudinal gradient in speciation rate for marine fishes. *Nature* 559, 392–395.
- Ramos, W.R., Medeiros, J.F., Julião, G.R., Ríos-Velásquez, C.M., Marialva, E.F., et al., 2014. Anthropogenic effects on sand fly (Diptera: psychodidae) abundance and diversity in an Amazonian rural settlement, Brazil. *Acta Trop.* 139, 44–52 <https://doi.org/10.1016/j.actatropica.2014.06.017>.
- Ready, P.D., 2013. Biology of phlebotomine sand flies as vectors of disease agents. *Annu. Rev. Entomol.* 58, 227–250.
- Restrepo Martínez, C., 2011. The karst system of La Danta (Sonson-Antioquia) Colombia. *Dyna (Medellin)* 78, 239–246.
- Rigg, C.A., Perea, M., González, K., Saldaña, A., Calzada, J.E., et al., 2021. Diversity, Co-Occurrence, and Nestedness Patterns of Sand Fly Species (Diptera: psychodidae) in Two Rural Areas of Western Panamá. *Insects* 12, 113. <https://doi.org/10.3390/insects12020113>.
- Romero Ricardo, L., Meza, N.L., Pérez Doria, A., Bejarano Martínez, E., 2013. *Lutzomyia abnenneci* and *Lutzomyia olmeca* (Diptera: psychodidae), new records for the department of Sucre. *Colombia. Acta Biológica Colombiana* 18, 375–380.
- Rutledge, L.C., Mosser, H.L., 1972. Biology of Immature Sandflies (Diptera: psychodidae) at the Bases of Trees in Panama 1. *Environ. Entomol.* 1, 300–309. <https://doi.org/10.1093/ee/1.3.300>.
- Santamaría, E., Cabrera, O.L., Marceló, C., Goenaga-Olaya, S., Maestre-Serrano, R., 2020. Behavioral aspects of phlebotomine sand flies associated with a case of cutaneous leishmaniasis in Atlántico, northern Colombia. *Am. J. Trop. Med. Hyg.* 102, 768.
- Santatana, A.L.F.D., Gpdy, R.E., Souza, N.A., Silva, J.D.S., Gómez, G.M., Palomino, N. R., Benavides, C.M., Lázaro, A.C., Guardo, M., Azevedo, A.C.R.D., 2020. Morphological Anomaly in the Scutum of *Bichromomyia olmeca bicolor* (Diptera: Psychodidae), Captured in Inapari Town, Trinitational Border Peru, Brazil and Bolivia. *Shannon, R.C., 1926. The occurrence of Phlebotomus in Panama. J. Washington Acad. Sci.* 16, 190–193.
- Shimabukuro, P.H.F., de Andrade, A.J., Galati, E.A.B., 2017. Checklist of American sand flies (Diptera, Psychodidae, Phlebotominae): genera, species, and their distribution. *Zookeys* (660), 67.
- Stuart-Smith, R.D., Bates, A.E., Lefcheck, J.S., Duffy, J.E., Baker, S.C., et al., 2013. Integrating abundance and functional traits reveals new global hotspots of fish diversity. *Nature* 501, 539–542.
- Teodoro, L.M., de Lima Carvalho, G.M., Campos, A.M., Cerqueira, R.F.V., Souza-Silva, M., Ferreira, R.L., Barata, R.A., 2021. Phlebotomine sand flies (Diptera, Psychodidae) from iron ore caves in the State of Pará, Brazil. *Subterranean Biology* 37, 27.
- Tesh, R.B., Peralta, P.H., Shope, R.E., Chaniotis, B.N., Johnson, K.M., 1975. Antigenic relationships among *Phlebotomus* fever group arboviruses and their implications for the epidemiology of sandfly fever. *Am. J. Trop. Med. Hyg.* 24, 135–144.
- Toprak, S., Özer, N., 2007. Distribution of Sand Fly (Diptera: psychodidae) Species and Efficiency of Capturing Methods in Sanliurfa Province. *Turkey. J. Med. Entomol.* 44, 23–28. <https://doi.org/10.1093/jmedent/41.5.23>.
- Vilela, M.L., Azevedo, A.C.R., Godoy, R.E., 2015. Description of a new phlebotomine species of the Brazilian Cerrado from sandstone caves in Tocantins State, Brazil: *Lutzomyia (Lutzomyia) elizabethrangela* sp. nov. (Diptera: psychodidae). *J. Med. Entomol.* 52, 596–603.
- Vivero, R.J., Muskus, C.E., Uribe, S.I., Bejarano, E.E., Torres, C., 2010. Flebotomíneos (Diptera: psychodidae) en la reserva natural del cañón del río Claro (Antioquia). *Colombia. Actualidades Biológicas* 32, 165–171.
- Vivero, R.J., Torres-Gutiérrez, C., Bejarano, E.E., Peña, H.C., Estrada, L.G., et al., 2015. Study on natural breeding sites of sand flies (Diptera: phlebotominae) in areas of *Leishmania* transmission in Colombia. *Parasit. Vectors* 8, 1–14.
- Wagner R., 2004 *Fauna Europaea: psychodidae*. *Fauna Europaea Web Service version 2*.
- Williams, P., 1976. The phlebotomine sandflies (Diptera, Psychodidae) of caves in Belize. *Central America. Bull. Entomol. Res.* 65, 601–614.
- Young, D.G., Arias, J.R., 1982. A new phlebotomine sand fly in the *Lutzomyia flaviscutellata* complex (Diptera: psychodidae) from Northern Brazil. *J. Med. Entomol.* 19, 134–138.
- Young, D.G., Duncan, M.A. 1994. Guide to the identification and geographic distribution of *Lutzomyia* sand flies in Mexico, the West Indies, Central and South America (Diptera: Psychodidae). *Memoirs of the American Entomological Institute* 54, 881 pp. [1994.03.31].