

# The risk of COVID-19 death is much greater and age-dependent with type I IFN autoantibodies

**Jeremy Manry** (✉ [jeremy.manry@inserm.fr](mailto:jeremy.manry@inserm.fr))

INSERM U1163 <https://orcid.org/0000-0001-5998-2051>

**Paul Bastard**

Laboratory of Human Genetics of Infectious Diseases, Necker Branch, INSERM U1163

**Adrian Gervais**

INSERM U1163

**Tom Le Voyer**

INSERM U1163

**Jérémie Rosain**

Laboratory of Human Genetics of Infectious Diseases, Necker Branch, INSERM <https://orcid.org/0000-0002-2822-161X>

**Quentin Philippot**

INSERM U1163

**Eleftherios Michailidis**

The Rockefeller University, Laboratory of Virology and Infectious Disease <https://orcid.org/0000-0002-9907-4346>

**Hans-Heinrich Hoffmann**

Laboratory of Virology and Infectious Disease, Rockefeller University

**Shohei Eto**

Department of Pediatrics, Graduate School of Biomedical and Health Sciences, Hiroshima University

**Marina Garcia-Prat**

Pediatric Infectious Diseases and Immunodeficiencies Unit, Hospital Universitari Vall d'Hebron, Vall d'Hebron Research Institute

**Lucy Bizien**

INSERM U1163

**Alba Parra-Martínez**

Pediatric Infectious Diseases and Immunodeficiencies Unit, Hospital Universitari Vall d'Hebron, Vall d'Hebron Research Institute

**Rui Yang**

St. Giles Laboratory of Human Genetics of Infectious Diseases, Rockefeller Branch, Rockefeller University

**Liis Haljasmägi**

University of Tartu

**Mélanie Migaud**

INSERM U1163

**Karita Särekannu**

Institute of Biomedicine and Translational Medicine, University of Tartu

**Julia Maslovskaja**

Institute of Biomedicine and Translational Medicine, University of Tartu

**Nicolas de Prost**

Hôpitaux Universitaires Henri Mondor <https://orcid.org/0000-0002-4833-4320>

**Yacine Tandjaoui-Lambiotte**

Avicenne Hospital, AP-HP, Bobigny, INSERM U1272, Hypoxia and Lung

**Charles-Edouard Luyt**

Hôpital Pitié-Salpêtrière, Service de Médecine Intensive Réanimation, Institut de Cardiologie

**Blanca Amador-Borrero**

Internal Medicine Department, Lariboisière Hospital AP-HP, Paris University

**Alexandre Gaudet**

University of Lille, U1019-UMR9017, Center for Infection and Immunity of Lille

**Julien Poissy**

University of Lille, U1019-UMR9017, Center for Infection and Immunity of Lille

**Pascal Morel**

Etablissement Français Du Sang

**Pascale Richard**

Etablissement Français Du Sang <https://orcid.org/0000-0003-1864-3824>

**Fabrice Cognasse**

Etablissement Français du Sang, Auvergne-Rhône-Alpes

**Jesus Troya**

Department of Internal Medicine, Infanta Leonor University Hospital

**Sophie Trouillet-Assant**

Hospices Civils de Lyon

**Alexandre Belot**

Hospices Civils de Lyon <https://orcid.org/0000-0003-4902-5332>

**Kahina Saker**

Hospices Civils de Lyon <https://orcid.org/0000-0001-8825-5400>

**Pierre Garçon**

Intensive Care Unit, Grand Hôpital de l'Est Francilien Site de Marne-La-Vallée

**Jacques G. Rivière**

Hospital Universitari Vall d'Hebron <https://orcid.org/0000-0003-1055-2063>

**Jean-Christophe Lagier**

Méditerranée Infection Foundation

**Stéphanie Gentile**

Service d'Evaluation Médicale, Hôpitaux Universitaires de Marseille APMH

**Lindsey Rosen**

National Institutes of Health

**Elana Shaw**

Laboratory of Clinical Immunology and Microbiology, National Institute of Allergy and Infectious Diseases, National Institutes of Health,

**Tomohiro Morio**

Tokyo Medical and Dental University <https://orcid.org/0000-0002-9259-1025>

**Junko Tanaka**

Department of Epidemiology, Infectious Disease Control and Prevention, Graduate School of Biomedical and Health Sciences, Hiroshima University

**David Dalmau**

Hospital Universitari MútuaTerrassa; Fundació Docència i Recerca MutuaTerrassa, Terrasa; Universitat de Barcelona

**Pierre-Louis Tharaux**

Institut National de la Santé et de la Recherche Médicale (INSERM) <https://orcid.org/0000-0002-6062-5905>

**Damien Sene**

Internal Medicine Department, Lariboisière Hospital AP-HP, Paris University

**Alain Stepanian**

Service d'Hématologie Biologique, Hôpital Lariboisière, AP-HP and EA3518, Institut Universitaire d'Hématologie-Hôpital Saint Louis, Université Paris

**Bruno Mégarbane**

Réanimation Médicale et Toxicologique, Hôpital Lariboisière (AP-HP), Université Paris-Diderot, INSERM Unité Mixte de Recherche Scientifique (UMRS) 1144

**Vasiliki Triantafyllia**

Laboratory of Immunobiology, Center for Clinical, Experimental Surgery, and Translational Research, Biomedical Research Foundation of the Academy of Athens

**Arnaud Fekkar**

INSERM U1163

**James Heath**

Institute for Systems Biology

**Jose Franco**

University of Antioquia <https://orcid.org/0000-0001-5664-6415>

**Juan-Manuel Anaya**

Universidad del Rosario <https://orcid.org/0000-0002-6444-1249>

**Jordi Solé-Violán**

Intensive Care Medicine, University Hospital of Gran Canaria Dr. Negrín, Canarian Health System

**Luisa Imberti**

CREA Laboratory (AIL Center for Hemato-Oncologic Research), Diagnostic Department, ASST Spedali Civili di Brescia

**Andrea Biondi**

Fondazione Tettamenti

**Paolo Bonfanti**

Department of Infectious Diseases, San Gerardo Hospital, University of Milano Bicocca

<https://orcid.org/0000-0001-7289-8823>

**Riccardo Castagnoli**

Laboratory of Clinical Immunology and Microbiology, National Institute of Allergy and Infectious Diseases, National Institutes of Health, <https://orcid.org/0000-0003-0029-9383>

**Ottavia Delmonte**

Immune Deficiency Genetics Section, Laboratory of Host Defenses, Division of Intramural Research, National Institute of Allergy and Infectious Diseases, National Institutes of Health

**Yu Zhang**

NIAID <https://orcid.org/0000-0002-6904-0372>

**Andrew Snow**

Uniformed Services University of the Health Sciences, Bethesda, MD <https://orcid.org/0000-0002-8728-6691>

**Steve Holland**

Division of Intramural Research (HNM2), National Institute of Allergy and Infectious Diseases

<https://orcid.org/0000-0003-3207-5464>

**Catherine Biggs**

Department of Pediatrics, British Columbia Children's Hospital, University of British Columbia

**Marcela Moncada-Vélez**

St. Giles Laboratory of Human Genetics of Infectious Diseases, Rockefeller Branch, Rockefeller University

**Andrés Arias**

St. Giles Laboratory of Human Genetics of Infectious Diseases, Rockefeller Branch, Rockefeller University

**Lazaro Lorenzo**

Necker Medical School

**Soraya Boucherit**

Necker Medical School <https://orcid.org/0000-0002-8819-7594>

**Dany Anglicheau**

CHU Necker

**Anna Planas**

Spanish National Research Council <https://orcid.org/0000-0002-6147-1880>

**Filomeen Haerynck**

Ghent University Hospital

**Sotirija Duvlis**

Faculty of Medical Sciences, University "Goce Delchev," Štip, Republic of Northern Macedonia.

<https://orcid.org/0000-0001-8587-7386>

**Robert Nussbaum**

Invitae (United States)

**Tayfun Ozcelik**

Bilkent University <https://orcid.org/0000-0001-5937-1082>

**Sevgi Keles**

Necmettin Erbakan University, Meram Medical Faculty <https://orcid.org/0000-0001-7344-8947>

**Aziz Bousfiha**

Centre Hospitalier Universitaire Hassan II

**Jalila El Bakkouri**

Clinical Immunology Unit, Department of Pediatric Infectious Disease, CHU Ibn Rushd and LICIA, Laboratoire d'Immunologie Clinique, Inflammation et Allergie, Faculty of Medicine and Pharmacy,

**Carolina Ramirez-Santana**

Center for Autoimmune Disease Research, School of Medicine and Health Sciences, Universidad del Rosario, Bogotá, Colombia.

**Stéphane Paul**

Centre International de Recherche en Infectiologie Lyon <https://orcid.org/0000-0002-8830-4273>

**Qiang Pan-Hammarstrom**

Karolinska Institute <https://orcid.org/0000-0003-1990-8804>

**Lennart Hammarstrom**

Karolinska Institute

**Annabelle Dupont**

Université de Lille, INSERM, CHU de Lille, Institut Pasteur de Lille, U1011-EGID, F-59000 Lille, France.

**Alina Kurolap**

Genetics Institute, Tel Aviv Sourasky Medical Center, Tel Aviv University, Tel Aviv, Israel.

**Christine Metz**

Feinstein Institute for Medical Research

**Alessandro Aiuti**

San Raffaele Telethon Institute for Gene Therapy (SR-Tiget), IRCCS San Raffaele Scientific Institute, Milan <https://orcid.org/0000-0002-5398-1717>

**Giorgio Casari**

Vita-Salute San Raffaele University, and Clinical Genomics, IRCCS Ospedale San Raffaele, Milan, Italy.

<https://orcid.org/0000-0002-0115-8980>

**Vito Lampasona**

IRCCS Ospedale San Raffaele <https://orcid.org/0000-0001-5162-8445>

**Fabio Ciceri**

Hematology and Bone Marrow Transplantation Unit, IRCCS San Raffaele Scientific Institute, Milano, Italy.

**Lucila Barreiros**

Department of Immunology, Institute of Biomedical Sciences, University of São Paulo, São Paulo, Brazil.

**Elena Dominguez-Garrido**

Fundación Rioja Salud, Centro de Investigación Biomédica de La Rioja, Logroño, Spain.

**Mateus Vidigal**

University of São Paulo, São Paulo, Brazil.

**Mayana Zatz**

University of São Paulo, São Paulo, Brazil.

**Diederik van de Beek**

Department of Neurology, Amsterdam Neuroscience, Amsterdam, Netherlands.

**Sabina Sahanic**

Department of Internal Medicine II, Medical University Innsbruck

**Ivan Tancevski**

Innsbruck Medical University

**Yurii Stepanovskyy**

Shupyk National Healthcare University of Ukraine, Kyiv, Ukraine.

**Oksana Boyarchuk**

Department of Children's Diseases and Pediatric Surgery, I. Horbachevsky Ternopil National Medical University, Ternopil, Ukraine.

**Yoko Nukui**

Department of Infection Control and Prevention, Medical Hospital, TMDU, Tokyo, Japan.

**Miyuki Tsumura**

Hirosima University

**Loreto Vidaur**

Intensive Care Medicine, Donostia University Hospital, Biodonostia Institute of Donostia, San Sebastián, Spain.

**Stuart Tangye**

Garvan Institute <https://orcid.org/0000-0002-5360-5180>

**Sonia Burrel**

Sorbonne University <https://orcid.org/0000-0002-7783-2601>

**Darragh Duffy**

Translational Immunology Lab, Institut Pasteur <https://orcid.org/0000-0002-8875-2308>

**Lluís Quintana-Murci**

Institut Pasteur <https://orcid.org/0000-0003-2429-6320>

**Adam Klocperk**

Department of Immunology, Second Faculty of Medicine, Charles University and University Hospital Motol, 15006 Prague

**Nelli Kann**

Dmitry Rogachev National Medical Research Center of Pediatric Hematology, Oncology and Immunology, Moscow, Russia.

**Anna Shcherbina**

Dmitry Rogachev National Medical Research Center of Pediatric Hematology, Oncology and Immunology, Moscow, Russia.

**Yu-Lung Lau**

The University of Hong Kong

**Daniel Leung**

Department of Paediatrics and Adolescent Medicine, University of Hong Kong, Hong Kong, China.

**Matthieu Coulangeat**

Division of Geriatric Medicine, Tours University Medical Center, Tours, France. <https://orcid.org/0000-0003-1986-3546>

**Julien Marlet**

INSERM U1259, MAVIVH, Université de Tours, Tours, France. <https://orcid.org/0000-0002-8645-8703>

**Rutger Koning**

Department of Neurology, Amsterdam Neuroscience, Amsterdam, Netherlands.

**Luis Reyes**

Department of Microbiology, Universidad de La Sabana, Chía, Colombia.

**Angélique Chauvineau-Grenier**

Service de Biologie Médicale, CHI Robert Ballanger, Aulnay-sous-Bois, France.

**Fabienne Venet**

Hospices Civils de Lyon <https://orcid.org/0000-0003-0462-4235>

**guillaume monneret**

Immunology department <https://orcid.org/0000-0002-9961-5739>

**Michel Nussenzweig**

Rockefeller University

**Romain Arrestier**

Service de Médecine Intensive Réanimation, Hôpitaux Universitaires Henri Mondor, Assistance Publique-Hôpitaux de Paris (AP-HP), Paris, France.

**Idris Boudhabhay**

Department of Nephrology and Transplantation, Necker University Hospital, APHP, Paris, France. 58INEM, INSERM U1151–CNRS UMR 8253, Paris University, Paris, France.

**Hagit Baris-Feldman**

Genetics Institute, Tel Aviv Sourasky Medical Center, Tel Aviv University, Tel Aviv, Israel.

**David Hagin**

Sackler Faculty of Medicine, Tel-Aviv University, Tel-Aviv

**Joost Wauters**

Medical Intensive Care Unit, UZ Gasthuisberg & Laboratory for Clinical Infectious and Inflammatory Disorders, Department of Microbiology, Immunology and Transplantation, KU Leuven  
<https://orcid.org/0000-0002-5983-3897>

**Isabelle Meyts**

University Hospitals Leuven <https://orcid.org/0000-0003-1214-0302>

**Adam Dyer**

Department of Age-Related Healthcare, Tallaght University Hospital, Dublin, Ireland.

**Sean Kennelly**

Department of Age-Related Healthcare, Tallaght University Hospital, Dublin, Ireland.

**Nollaig Bourke**

Department of Medical Gerontology, School of Medicine, Trinity College Dublin, Dublin, Ireland.

**Rabih Halwani**

SIMR <https://orcid.org/0000-0002-6516-7771>

**Fatemeh Sharif-Askari**

Sharjah Institute for Medical Research, College of Medicine, University of Sharjah, Sharjah, United Arab Emirates.

**Karim Dorgham**

Sorbonne Universités, UPMC Univ Paris 06, INSERM, CNRS, Centre d'Immunologie et des Maladies Infectieuses (CIMIParis UMRS 1135) <https://orcid.org/0000-0001-9539-3203>

**Jérôme Sallette**

Cerba HealthCare, Issy-les-Moulineaux, France.

**Souad Mehlal-Sedkaoui**

Cerba HealthCare, Issy-les-Moulineaux, France.

**Suzan AlKhater**

Department of Pediatrics, King Fahad Hospital of the University, Al Khobar, Saudi Arabia.

**Raúl Rigo-Bonnin**

Department of Clinical Laboratory, Hospital Universitari de Bellvitge, IDIBELL, Barcelona, Spain.

**Francisco Morandeira**

Department of Immunology, Hospital Universitari de Bellvitge, IDIBELL, Barcelona, Spain.

**Lucie Roussel**

Department of Medicine, Division of Infectious Diseases, McGill University Health Centre, Montréal, QC, Canada.

**Donald Vinh**

The Research Institute of the McGill University Health Centre <https://orcid.org/0000-0003-1347-7767>

**Christian Erikstrup**

Aarhus University Hospital <https://orcid.org/0000-0001-6551-6647>

**Antonio Condino-Neto**

Institute of Biomedical Sciences, University of São Paulo

**Carolina Prando**



Faculdades Pequeno Príncipe, Instituto de Pesquisa Pelé Pequeno Príncipe, Curitiba, Brazil.

<https://orcid.org/0000-0002-9570-9770>

**Anastasiia Bondarenko**

Shupyk National Healthcare University of Ukraine, Kyiv, Ukraine.

**András Spaan**

St. Giles Laboratory of Human Genetics of Infectious Diseases, Rockefeller Branch, Rockefeller University, New York, NY, USA.

**Laurent Gilardin**

Service de Médecine Interne, Hôpital universitaire Jean-Verdier AP-HP, Bondy, France.

**Jacques Fellay**

École Polytechnique Fédérale de Lausanne <https://orcid.org/0000-0002-8240-939X>

**Stanislas Lyonnet**

Hôpital Necker-Enfants Malades

**Kaya Bilguvar**

Yale University School of Medicine

**Richard Lifton**

Laboratory of Human Genetics and Genomics, The Rockefeller University

**Shrikant Mane**

Yale University School of Medicine

**Mark Anderson**

Diabetes Center, University of California San Francisco, San Francisco, CA, USA.

**Bertrand Boisson**

Rockefeller University

**Vivien Béziat**

INSERM U1163 <https://orcid.org/0000-0002-4020-824X>

**Shen-Ying Zhang**

<https://orcid.org/0000-0002-9449-3672>

**Evangelos Andreakos**

Biomedical Research Foundation, Academy of Athens <https://orcid.org/0000-0001-5536-1661>

**Olivier Hermine**

Institut National de la Santé et de la Recherche Médicale, Unité Mixte de Recherche (UMR) 1163

**Aurora Pujol**

ICREA/ IDIBELL <https://orcid.org/0000-0002-9606-0600>

**Pärt Peterson**

Molecular Pathology Research Group, Institute of Biomedicine and Translational Medicine, University of Tartu <https://orcid.org/0000-0001-6755-791X>

**Trine Hyrup Mogensen**

Aarhus University <https://orcid.org/0000-0002-1853-9704>

**Lee Rowen**

Institute for Systems Biology, Seattle, WA, USA.

**James Mond**

ADMA Biologics Inc., Ramsey, NJ, USA

**Stéphanie Debette**

University of Bordeaux, Inserm, Bordeaux Population Health Research Center, UMR 1219

<https://orcid.org/0000-0001-8675-7968>

**Xavier deLamballerie**

Aix-Marseille University

**Charles Burdet**

INSERM CIC 1425, Paris, France.

**Lila Bouadma**

APHP- Hôpital Bichat – Médecine Intensive et Réanimation des Maladies

**Marie Zins**

Université de Paris, Université Paris-Saclay, UVSQ, INSERM UMS11, Villejuif, France.

<https://orcid.org/0000-0002-4540-4282>

**Pere Soler-Palacin**

Vall d'Hebron University Hospital

**Roger Colobran**

Hospital Universitari Vall d'Hebron (HUVH) <https://orcid.org/0000-0002-5964-536X>

**Guy Gorochov**

APHP Sorbonne universite <https://orcid.org/0000-0003-2097-9677>

**Xavier Solanich**

Department of Internal Medicine, Hospital Universitari de Bellvitge, IDIBELL, Barcelona, Spain.

**Sophie Susen**

Université de Lille, INSERM, CHU de Lille, Institut Pasteur de Lille, U1011-EGID, F-59000 Lille, France.

**Javier Martinez-Picado**

IrsiCaixa <https://orcid.org/0000-0002-4916-2129>

**Didier Raoult**

Aix Marseille Université; IHU Méditerranée Infection-MEPHI <https://orcid.org/0000-0002-0633-5974>

**Marc Vasse**

Service de Biologie Clinique and UMR-S 1176, Hôpital Foch, Suresnes, France.

**Peter Gregersen**

Feinstein Institutes for Medical Research, Northwell Health, Manhasset, NY, USA.

**Carlos Rodríguez-Gallego**

Department of Immunology, University Hospital of Gran Canaria Dr. Negrin, Canarian Health System, Las Palmas de Gran Canaria, Spain.

**Lorenzo Piemonti**

IRCCS Ospedale San Raffaele , San Raffaele Diabetes Research Institute, Via Olgettina 60, 20132 Milan

<https://orcid.org/0000-0002-2172-2198>

**Luigi Notarangelo**

NIAID/NIH <https://orcid.org/0000-0002-8335-0262>

**Helen Su**

NIAID, NIH <https://orcid.org/0000-0002-5582-9110>

**Kai Kisand**

University of Tartu <https://orcid.org/0000-0002-5426-4648>

**Satoshi Okada**

Hiroshima University Graduate School of Biomedical and Health Sciences <https://orcid.org/0000-0002-4622-5657>

**Anne Puel**

INSERM

**Emmanuelle Jouanguy**

Rockefeller University

**Charles Rice**

Rockefeller University <https://orcid.org/0000-0003-3087-8079>

**Pierre Tiberghien**

EFS

**Qian Zhang**

St. Giles Laboratory of Human Genetics of Infectious Diseases, Rockefeller Branch, Rockefeller University, New York, NY, USA.

**Jean-Laurent Casanova****Laurent Abel**

Necker Medical School

**Aurélie Cobat**

INSERM <https://orcid.org/0000-0001-7209-6257>

---

**Article****Keywords:**

**Posted Date:** January 14th, 2022

**DOI:** <https://doi.org/10.21203/rs.3.rs-1225906/v1>

**License:**  This work is licensed under a Creative Commons Attribution 4.0 International License.

[Read Full License](#)

---

**Version of Record:** A version of this preprint was published at Proceedings of the National Academy of Sciences on May 16th, 2022. See the published version at <https://doi.org/10.1073/pnas.2200413119>.

# Abstract

SARS-CoV-2 infection fatality rate (IFR) doubles with every five years of age from childhood onward. Circulating autoantibodies neutralizing IFN- $\alpha$ , IFN- $\omega$ , and/or IFN- $\beta$  are found in ~20% of deceased patients across age groups. In the general population, they are found in ~1% of individuals aged 20-70 years and in >4% of those >70 years old. With a sample of 1,261 deceased patients and 34,159 uninfected individuals, we estimated both IFR and relative risk of death (RRD) across age groups for individuals carrying autoantibodies neutralizing type I IFNs, relative to non-carriers. For autoantibodies neutralizing IFN- $\alpha$ 2 or IFN- $\omega$ , the RRD was 17.0[95% CI:11.7-24.7] for individuals under 70 years old and 5.8[4.5-7.4] for individuals aged 70 and over, whereas, for autoantibodies neutralizing both molecules, the RRD was 188.3[44.8-774.4] and 7.2[5.0-10.3], respectively. IFRs increased with age, from 0.17%[0.12-0.31] for individuals <40 years old to 26.7%[20.3-35.2] for those  $\geq$ 80 years old for autoantibodies neutralizing IFN- $\alpha$ 2 or IFN- $\omega$ , and from 0.84%[0.31-8.28] to 40.5%[27.82-61.20] for the same two age groups, for autoantibodies neutralizing both molecules. Autoantibodies against type I IFNs increase IFRs, and are associated with high RRDs, particularly those neutralizing both IFN- $\alpha$ 2 and - $\omega$ . Remarkably, IFR increases with age, whereas RRD decreases with age. Autoimmunity to type I IFNs appears to be second only to age among common predictors of COVID-19 death.

## Introduction

There have already been more than 250 million SARS-CoV-2 infections and at least five million deaths from COVID-19 worldwide. Interindividual clinical variability in the course of infection with SARS-CoV-2 is immense, ranging from silent infection in about 40% of cases to acute respiratory distress syndrome in ~3% of cases<sup>1-3</sup>. Death occurs in ~1% of cases<sup>4</sup>. Age is the strongest epidemiological predictor of COVID-19 death, with the risk of death doubling every five years of age from childhood onward<sup>4,5</sup>. Men are also at greater risk of death than women<sup>3,6</sup>. The COVID Human Genetic Effort<sup>7</sup> has shown that type I interferon (IFN) immunity is essential for protective immunity to respiratory infection with SARS-CoV-2<sup>8-11</sup>. We have reported that inborn errors of Toll-like receptor 3 (TLR3)-dependent type I IFN immunity can underlie life-threatening COVID-19 pneumonia in a small subset of patients<sup>11</sup>. Biochemically deleterious mutations of eight genes were found in 23 patients with critical COVID-19 (3.5% of 659 patients), including 18 patients under 60 years old. Remarkably, four patients, aged 25 to 50 years, had autosomal recessive (AR) deficiency of IFNAR1 or IRF7. Two other patients with AR IFNAR1 or TBK1 deficiency were independently reported<sup>12,13</sup>. The penetrance of those defects is unknown, but it is probably higher for AR than for autosomal dominant disorders. We then reported that X-linked recessive TLR7 deficiency accounted for 1.8% of cases of life-threatening COVID-19 in men under 60 years old<sup>10,14</sup>. The penetrance of this disorder is apparently high but incomplete, especially in children. Deficiencies of IFNAR1 and IRF7 blunt type I IFN immunity across cell types, whereas defects of the TLR3 and TLR7 pathway preferentially affect respiratory epithelial cells and plasmacytoid dendritic cells, respectively<sup>10,15</sup>.

We have also reported the presence of autoantibodies (auto-Abs) neutralizing high concentrations (10 ng/mL, with plasma diluted 1/10) of IFN- $\alpha$ 2 and/or IFN- $\omega$  in about 10% of patients with critical COVID-19 pneumonia but not in individuals with asymptomatic or mild infection<sup>9</sup>. This finding has already been replicated in 13 other cohorts<sup>16-29</sup>. We then detected auto-Abs neutralizing lower, more physiological concentrations (100 pg/mL, with plasma diluted 1/10) of IFN- $\alpha$ 2 and/or IFN- $\omega$  in 13.6% of patients with life-threatening COVID-19, and 18% of deceased patients<sup>8</sup>. The proportion of male patients was greater in patients with auto-Abs than in patients without auto-Abs<sup>8,9</sup>. In addition, 1.3% of patients with critical COVID-19 had auto-Abs neutralizing IFN- $\beta$  (10 ng/mL, with plasma diluted 1/10), most without auto-Abs neutralizing IFN- $\alpha$ 2 or IFN- $\omega$ . The prevalence of auto-Abs neutralizing IFN- $\alpha$ 2 and/or IFN- $\omega$  in the general population increased with age, from 0.18% for 10 ng/mL and 1% for 100 pg/mL in individuals between 18 and 69 years old to 3.4% for 10 ng/mL and 6.3% for 100 pg/mL for individuals over 80 years old<sup>8</sup>. The prevalence of auto-Abs against IFN- $\beta$  did not increase with age. The crude odds ratios (ORs) for critical COVID-19 as opposed to asymptomatic or mild infection in auto-Ab carriers relative to non-carriers ranged from 3 to 67, depending on the type I IFNs recognized and the concentrations neutralized<sup>8</sup>. At least 12 lines of evidence strongly suggest that auto-Abs against type I IFNs are strong determinants of COVID-19 death (Table 1). The specific impact of these auto-Abs on COVID-19 mortality according to age and sex remains unknown and is of major interest, as both the prevalence of these auto-Abs and the risk of death increase with age and are higher in men. Here, we estimated the relative risk of COVID-19 death (RRD) and the SARS-CoV-2 infection fatality rate (IFR) for type I IFN auto-Ab carriers relative to non-carriers, by sex and age category.

## Methods

### Study design

We enrolled 1,261 patients aged 20 to 99 years old who died from COVID-19 pneumonia, and 34,159 controls from the adult general population from whom samples were collected before the COVID-19 pandemic, as previously described<sup>8</sup>. All subjects were recruited according to protocols approved by local institutional review boards (IRBs). Auto-Ab determinations were performed as described by Bastard *et al.*<sup>8,46</sup>, and were classified as neutralizing high concentrations (10 ng/ml) of IFN- $\alpha$ 2, - $\omega$ , or - $\beta$ , or low concentrations (100 pg/ml) of IFN- $\alpha$ 2, or - $\omega$  (Additional Methods).

### RRDs and IFRs for carriers of neutralizing autoantibodies

We estimated the RRD in individuals carrying auto-Abs neutralizing type I IFNs relative to non-carriers, using large samples of patients who died from COVID-19 and of individuals from the general population. For each combination of auto-Abs, a Firth's bias-corrected logistic regression model, including auto-Ab status, sex and age was fitted (Supplementary Table 1). For assessments of the effect of age and sex on the RRD due to auto-Abs, we added auto-Abs\*sex and auto-Abs\*age interaction terms to the Firth's logistic regression model (Additional Methods). We estimated the IFR for carriers of neutralizing auto-Abs

infected with SARS-CoV-2 ( $IFR_{AAB}$ ) following Bayes' theorem, and using the age-dependent prevalence of auto-Abs in deceased patients and in the general population together with the reported age-specific  $IFR^4$  as detailed in Additional Methods.

## Results

### Patients and controls

We estimated the RRD of individuals carrying auto-Abs neutralizing type I IFNs relative to non-carriers by Firth's logistic regression, using large samples of 1,261 patients who died from COVID-19 and 34,159 individuals from the general population from whom samples were collected before the pandemic. In this study design, in which controls are sampled from the baseline population regardless of disease status, the ORs obtained by logistic regression approximate to the RR in the absence of the assumption of rare disease<sup>47</sup> (see Additional Methods). For auto-Abs neutralizing low concentrations (100 pg/mL) of IFN- $\alpha$ 2 and/or IFN- $\omega$ , we used 1,121 patients who died from COVID-19, and 10,778 individuals from the general population (Table 2).

Assessments of auto-Abs neutralizing high concentrations (10 ng/mL) of IFN- $\alpha$ 2 and/or IFN- $\omega$  were available for 1,094 deceased patients, and 34,159 individuals from the general population (Table 2). We also had assessments of auto-Abs neutralizing 10 ng/mL IFN- $\beta$  for a subsample of 636 deceased patients and 9,126 individuals from the general population (Table 2). RRD was estimated by means of Firth's bias-corrected logistic regression, considering death as a binary outcome and adjusting for sex and age in six classes (20-39, 40-49, 50-59, 60-69, 70-79,  $\geq 80$  years). For assessment of the effect of age and sex on RRD, we added auto-Abs\*age and auto-Abs\*sex interaction terms to the logistic model (see Methods and Additional Methods).

### RRD for carriers of auto-Abs neutralizing low concentrations of type I IFNs

We first estimated the RRD for individuals carrying auto-Abs neutralizing low concentrations of IFN- $\alpha$ 2 or IFN- $\omega$ . As expected, increasing age and maleness were highly significantly associated with greater risk of COVID-19 death ( $P$  values  $\leq 10^{-16}$ , Supplementary Table 1). Different age classes were used to test the interaction with the presence of auto-Abs, and the best fit was obtained with a two-age class model (20-69 and  $\geq 70$  years, Supplementary Table 2) with a significant effect of the auto-Abs\*age interaction term ( $P$  value =  $4 \times 10^{-6}$ ). The RRD associated with auto-Abs did not vary significantly with sex ( $P$  value = 0.81). These interaction results are fully consistent with the distribution of RRD according to age (Fig. 1A) and sex (Fig. 1B), with a clear decrease in RRD after the age of 70 years, and no sex effect. Overall, the RRD for individuals carrying auto-Abs neutralizing IFN- $\alpha$ 2 or IFN- $\omega$  decreased from 17.0 [95% CI: 11.7-24.7] before the age of 70 years to 5.8 [4.5-7.4] for individuals  $\geq 70$  years old (Fig. 2A, Supplementary Table 3). We then applied the same strategy to other combinations of auto-Abs neutralizing low concentrations of

IFN, and observed similar age effects on RRDs (Supplementary Table 1). The presence of auto-Abs neutralizing both IFN- $\alpha$ 2 and IFN- $\omega$  was associated with the highest RRD, estimated at 188.3 [45.8-774.4] for individuals under the age of 70 years and 7.2 [5.0-10.3] for those over 70 years old (Fig. 2A, Supplementary Table 3).

### **RRD for carriers of auto-Abs neutralizing high concentrations of type I IFNs**

We then estimated the RRD for the presence *versus* the absence of auto-Abs neutralizing high concentrations (10 ng/mL) of type I IFN. The effect of age on RRD was similar to that observed with auto-Abs neutralizing low concentrations of type I IFN, with the use of two age classes providing the best fit (Supplementary Table 2 and 4). The RRDs associated with auto-Abs neutralizing high concentrations of type I IFNs were higher than those associated with auto-Abs neutralizing low concentrations, and also decreased with age (Fig. 2B, Supplementary Table 5). The RRD for carriers of IFN- $\alpha$ 2 or IFN- $\omega$  auto-Abs decreased from 62.4 [38.4-101.3] before the age of 70 years to 6.8 [5.1-9.2] after the age of 70 years, whereas carriers of auto-Abs against both IFN- $\alpha$ 2 and IFN- $\omega$  had the highest RRD, estimated at 156.5 [57.8-423.4] and 12.9 [8.4-19.9] for subjects <70 years and  $\geq$ 70 years old, respectively (Fig. 2B, Supplementary Table 5). Interestingly, auto-Abs neutralizing high doses of IFN- $\beta$  had the lowest RRD before 70 years (7.0 [2.2-22.4]), with no significant age-dependent association ( $P$  value = 0.37).

### **IFR in individuals carrying auto-Abs neutralizing low concentrations of type I IFNs**

We then estimated the IFR in SARS-CoV-2-infected individuals carrying auto-Abs neutralizing low concentrations of type I IFNs. According to Bayes' theorem,  $IFR_{AAB}$  can be expressed as a function of the age-dependent prevalence of auto-Abs in deceased patients and in the general population together with the reported age-specific IFR<sup>4</sup> (see Supplementary Information). For all combinations of auto-Abs, the  $IFR_{AAB}$  was much higher than the overall IFR. Figure 3 illustrates this much higher IFR for carriers of auto-Abs neutralizing low concentrations of IFN- $\alpha$ 2 or IFN- $\omega$ ; it exceeded 1% and 10% for subjects over the ages of 40 and 60 years, respectively. Considering other combinations of auto-Abs, the highest  $IFR_{AAB}$  was observed for carriers of auto-Abs neutralizing both IFN- $\alpha$ 2 and - $\omega$ , reaching 40.5% [27.8-61.2] in individuals over 80 years old (Fig. 4A and Supplementary Table 6).  $IFR_{AAB}$  values were similar for all other combinations of auto-Abs. For example, the  $IFR_{AAB}$  for individuals carrying auto-Abs neutralizing either IFN- $\alpha$ 2 or - $\omega$  ranged from 0.17% [0.12-0.31] in individuals under 40 years old to 26.7% [20.3-35.2] in individuals over 80 years old. An exception was noted for the  $IFR_{AAB}$  of carriers of anti-IFN- $\alpha$ 2 auto-Abs, which was 1.8 to 2.6 times higher than that for carriers of auto-Abs neutralizing IFN- $\alpha$ 2 or - $\omega$  in subjects under 60 years old.. The  $IFR_{AAB}$  was also generally higher in male subjects than in female subjects, particularly in individuals carrying auto-Abs neutralizing both IFN- $\alpha$ 2 and - $\omega$  (~2.7 times higher) (Supplementary Fig. 1).

## IFR in individuals carrying auto-Abs neutralizing high concentrations of type I IFNs

The age-, sex- and type I IFN-dependent patterns of  $IFR_{AAB}$  observed for carriers of auto-Abs neutralizing high concentrations of IFN- $\alpha$ 2 and/or - $\omega$  were similar to those previously obtained for carriers of auto-Abs neutralizing low concentrations of these molecules, but with higher values. For example,  $IFR_{AAB}$  ranged from 3.1% [1.3-20.8] before 40 years of age to 68.7% [42.5-95.8] in those over 80 years old for carriers of auto-Abs neutralizing high concentrations of both IFN- $\alpha$ 2 and - $\omega$  (Fig. 4B, Supplementary Table 7).  $IFR_{AAB}$  values were ~5 times higher in male than in female subjects, across all age groups and auto-Abs combinations (Fig. 2). For carriers of auto-Abs neutralizing IFN- $\beta$  (tested only at high concentration),  $IFR_{AAB}$  was lower (by a factor of six to 71) than for individuals under the age of 80 years with auto-Abs neutralizing IFN- $\alpha$ 2 and/or - $\omega$ . It ranged from 0.04% [0.01-0.16] for individuals under the age of 40 years to 2.2% [0.2-9.3] for the 70-79 years age group. In the oldest age-class,  $IFR_{AAB}$  was 31.0% [2.4-88.1], similar to that for carriers of auto-Abs against IFN- $\alpha$ 2 or - $\omega$ , albeit with a large confidence interval.

## Discussion

In this study, we estimated RR from the general population<sup>47</sup> to obtain the RRDs associated with auto-Abs. We also used IFR values previously reported for the general population<sup>4</sup> to estimate  $IFR_{AAB}$ . We report high RRDs for carriers of auto-Abs neutralizing type I IFNs, ranging from 2.6 for auto-Abs neutralizing IFN- $\beta$  (high concentration) in subjects over 70 years old to >150 for auto-Abs neutralizing both IFN- $\alpha$ 2 and IFN- $\omega$  in subjects under 70 years old. For all types of auto-Abs, RRDs were three to 26 times higher in subjects under 70 years old than in older individuals. This is consistent with the increasing prevalence of auto-Abs in the general population with age (~1% under 70 years of age and >4% over 70 years of age), whereas the proportion of deceased patients with these auto-Abs is stable across age categories (~15-20%). The lower RRD observed in the elderly may be partly explained epidemiologically, by the larger contribution of other mortality risk factors, such as comorbid conditions, which become more frequent with increasing age. At the cellular level, aging is associated with immunosenescence, which may contribute to a defective innate and adaptive response to SARS-CoV-2 infection, thereby conferring a predisposition to severe COVID-19<sup>48</sup>. At the molecular level, global type I IFN immunity in the blood (plasmacytoid dendritic cells) and respiratory tract (respiratory epithelial cells) has been shown to decline with age<sup>49-52</sup>. These epidemiological, cellular, and molecular factors probably overlap. Thus, despite their increasing prevalence with age, auto-Abs against type I IFNs make a decreasing contribution to the risk of COVID-19 death with age due to the progressive development of additional age-dependent risk factors, including other mechanisms of type I IFN deficiency. However, for the very same reasons,  $IFR_{AAB}$  increases dramatically with age in patients with auto-Abs, reaching 68.7% for carriers of auto-Abs neutralizing high concentrations of both IFN- $\alpha$ 2 and - $\omega$ .



RRD and  $IFR_{AAB}$  varied considerably with the IFNs recognized and the concentrations neutralized by auto-Abs. For most combinations involving auto-Abs against IFN- $\alpha 2$  and/or  $-\omega$ , the neutralization of low concentrations was associated with a lower RRD and a lower  $IFR_{AAB}$  than the neutralization of high concentrations, suggesting that residual type I IFN activity may be beneficial in at least some patients. Blood IFN- $\alpha$  concentrations during acute asymptomatic or paucisymptomatic SARS-CoV-2 infection typically range from 1 to 100 pg/mL<sup>8</sup>. In addition, the presence of auto-Abs neutralizing both IFN- $\alpha 2$  and IFN- $\omega$  was associated with the highest RRD and  $IFR_{AAB}$  values. Interestingly, IFN- $\alpha 2$  and IFN- $\omega$  are encoded by two genes, *IFNA2* and *IFNW1*, that have been shown to have evolved under strong selective constraints<sup>53</sup>, consistent with their neutralization being harmful to the host. In addition, patients with auto-Abs against IFN- $\alpha 2$  have been shown to neutralize all 13 IFN- $\alpha$  subtypes<sup>8,9</sup>, rendering any potential IFN- $\alpha$  redundancy inoperative<sup>8,9</sup>. Accordingly, the  $IFR_{AAB}$  values for carriers of auto-Abs against IFN- $\alpha 2$  were higher than those for carriers of auto-Abs against IFN- $\omega$  in subjects under 60 years of age. In older age groups, this difference tended to disappear, consistent with the lower impact of auto-Abs in the elderly, as discussed above. Finally, auto-Abs neutralizing IFN- $\beta$  were less common, and associated with lower RRD and  $IFR_{AAB}$  values (by about one order of magnitude) than auto-Abs against IFN- $\alpha 2$  and/or IFN- $\omega$ , in all age groups except the over-80s. This less deleterious effect of auto-Abs neutralizing IFN- $\beta$  is consistent with a mouse study showing that the blockade of IFN- $\beta$  alone does not alter the early dissemination of lymphocytic choriomeningitis virus<sup>54</sup>. Overall, auto-Abs against type I IFNs are associated with very high RRD and IFR values, and the magnitude of this effect is much larger than that of other known common risk factors apart from age, such as maleness (Fig. 4), comorbidities, or the most significant common genetic variant on chromosome 3, all of which have been associated to life-threatening COVID-19 with ORs of about 2<sup>3</sup>.

Despite the lower prevalence of these auto-Abs in younger than in older individuals, the much higher  $IFR_{AAB}$  observed in individuals with these auto-Abs suggests that the testing of infected individuals in all age groups is warranted. Particular attention should be paid to patients, especially children, with known autoimmune or genetic conditions associated with the production of auto-Abs against type I IFNs. Early treatments could be provided<sup>55</sup>, including monoclonal antibodies<sup>56</sup>, new antiviral drugs, and/or IFN- $\beta$  in the absence of auto-Abs against IFN- $\beta$ <sup>57,58</sup>. Rescue treatment by plasma exchange is a therapeutic option in patients who already have pneumonia<sup>30</sup>. A screening of uninfected elderly people could be considered, given that these auto-Abs are found in 4% of individuals over 70 years old. Carriers of auto-Abs should be vaccinated against SARS-CoV-2 as a priority, and should benefit from a booster, whatever their age, and ideally from a monitoring of their antibody response to the vaccine. They should not receive live-attenuated vaccines, including the yellow fever vaccine (YFV-17D) and anti-SARS-CoV-2 vaccines based on the YFV-17D backbone<sup>46</sup>. In cases of SARS-CoV-2 infection, vaccinated patients should be closely monitored. As SARS-CoV-2 vaccination coverage increases and mortality due to COVID-19 decreases over time, it will be important to re-evaluate the risk of fatal COVID-19 in vaccinated individuals with and without auto-Abs. It is currently unclear whether these auto-Abs impair antibody responses to vaccines, and whether a vaccine-triggered antibody response can overcome type I IFN deficiency in response to

large or even medium-sized viral inocula. Finally, further investigations are required to determine the contribution of these auto-Abs to other severe viral diseases, and to elucidate the mechanisms underlying their development, which may be age-dependent. In the meantime, auto-Abs against type I IFNs should be considered as a leading common predictor of life-threatening COVID-19 after age, as their detection has a much greater predictive value for death, and, by inference, hospitalization and critical COVID-19, than sex, comorbidities, and common genetic variants (Fig. 3).

## Declarations

## Acknowledgments

We thank the patients and their families for placing their trust in us. We thank the members of both branches of the Laboratory of Human Genetics of Infectious Diseases. We thank Y. Nemirovskaya, M. Woollett, D. Liu, S. Boucherit, C. Rivalain, M. Chrabieh, and L. Lorenzo for administrative assistance. We also thank the staff of the Imagine facilities: C. Bureau, L. Colonna, S. Paillet, N. Ghouas, and M. Sy. We are also grateful to the legal team and technology transfer staff of the Imagine Institute: M. Pilorges, R. Marlanges, E. Rubino, W. Loewen, D. Beudin, and N. Wuytens. We thank all the staff of the Imagine Institute, Necker Hospital, and Necker sorting center for help. We thank S. Nagashima (Department of Epidemiology, Infectious Disease Control and Prevention, Graduate School of Biomedical and Health Sciences, Hiroshima University, Hiroshima, Japan).

## Funding/Support

The Laboratory of Human Genetics of Infectious Diseases is supported by the Howard Hughes Medical Institute, the Rockefeller University, the St. Giles Foundation, the National Institutes of Health (NIH) (R01AI088364 and R01AI163029), the National Center for Advancing Translational Sciences (NCATS), NIH Clinical and Translational Science Awards (CTSA) program (UL1 TR001866), a Fast Grant from Emergent Ventures, Mercatus Center at George Mason University, the Yale Center for Mendelian Genomics and the GSP Coordinating Center funded by the National Human Genome Research Institute (NHGRI) (UM1HG006504 and U24HG008956), the Yale High Performance Computing Center (S100D018521), the Fisher Center for Alzheimer's Research Foundation, the Meyer Foundation, the JPB Foundation, the French National Research Agency (ANR) under the "Investments for the Future" program (ANR-10-IAHU-01), the Integrative Biology of Emerging Infectious Diseases Laboratory of Excellence (ANR-10-LABX-62-IBEID), the French Foundation for Medical Research (FRM) (EQU201903007798), ANRS Nord-Sud (ANRS-COV05), ANR GENVIR (ANR-20-CE93-003), ANR AABIFNCOV (ANR-20-CO11-0001), ANR CNSVIRGEN (ANR-19-CE15-0009-01), and ANR GenMIS-C (ANR-21-COVR-0039) projects, the European Union's Horizon 2020 research and innovation program under grant agreement no. 824110 (EASI-Genomics), the Square Foundation, Grandir-Fonds de solidarité pour l'Enfance, the Fondation du Souffle, the SCOR Corporate

Foundation for Science, Institut National de la Santé et de la Recherche Médicale (INSERM), REACTing-INSERM; and the University of Paris. P.B. was supported by the FRM (EA20170638020). P.B., J.R., and T.L.V. were supported by the MD-PhD program of the Imagine Institute (with the support of Fondation Bettencourt Schueller). Work in the Laboratory of Virology and Infectious Disease was supported by the NIH (P01AI138398-S1, 2U19AI111825, and R01AI091707-10S1), a George Mason University Fast Grant, and the G. Harold and Leila Y. Mathers Charitable Foundation. The French COVID Cohort study group was sponsored by INSERM and supported by the REACTing consortium and by a grant from the French Ministry of Health (PHRC 20-0424). The Cov-Contact Cohort was supported by the REACTing consortium, the French Ministry of Health, and the European Commission (RECOVER WP 6). This work was also partly supported by the Intramural Research Program of the NIAID and NIDCR, NIH (grants ZIA AI001270 to L.D.N. and 1ZIAAI001265 to H.C.S.). This program is supported by the Agence Nationale de la Recherche (reference ANR-10-LABX-69-01). K.K.'s group was supported by the Estonian Research Council, through grants PRG117 and PRG377. R.H. was supported by an Al Jalila Foundation Seed Grant (AJF202019), Dubai, UAE, and a COVID-19 research grant (CoV19-0307) from the University of Sharjah, UAE. S.G.T. is supported by Investigator and Program Grants awarded by the National Health and Medical Research Council of Australia and a UNSW Sydney COVID Rapid Response Initiative Grant. L.I. and G.L.M reported funding from Regione Lombardia, Italy (project "Risposta immune in pazienti con COVID-19 e co-morbidità"). This research was partially supported by the Instituto de Salud Carlos III (COV20/0968). J.R.H. reported funding from Biomedical Advanced Research and Development Authority HHSO10201600031C. S.O. reports funding Research Program on Emerging and Re-emerging Infectious Diseases from Japan Agency for Medical Research and Development, AMED (grant number JP20fk0108531). G.G. was supported by ANR Flash COVID-19 program and SARS-CoV-2 Program of the Faculty of Medicine from Sorbonne University iCOVID programs. The Three-City (3C) Study was conducted under a partnership agreement between INSERM, Victor Segalen Bordeaux 2 University, and Sanofi-Aventis. The Fondation pour la Recherche Médicale funded the preparation and initiation of the study. The 3C Study was also supported by the Caisse Nationale d'Assurance Maladie des Travailleurs Salariés, Direction générale de la Santé, Mutuelle Générale de l'Education Nationale (MGEN), Institut de la Longévité, Conseils Régionaux of Aquitaine and Bourgogne, Fondation de France, and Ministry of Research-INSERM Program "Cohortes et collections de données biologiques". S. Debette was supported by the University of Bordeaux Initiative of Excellence. P.K.G. reports funding from the National Cancer Institute, NIH, under contract no. 75N91019D00024, task order no. 75N91021F00001. J.W. is supported by an FWO Fundamental Clinical Mandate (1833317N). Sample processing at IrsiCaixa was possible thanks to the crowdfunding initiative YoMeCorono. Work at Vall d'Hebron was also partly supported by research funding from Instituto de Salud Carlos III grant PI17/00660 cofinanced by the European Regional Development Fund (ERDF). C.R.-G. and colleagues from the Canarian Health System Sequencing Hub were supported by the Instituto de Salud Carlos III (COV20\_01333 and COV20\_01334, Spanish Ministry for Science and Innovation RTC-2017-6471-1; AEI/FEDER, UE), Fundación DISA (OA18/017 and OA20/024), and Cabildo Insular de Tenerife (CGIEU0000219140 and "Apuestas científicas del ITER para colaborar en la lucha contra la COVID-19"). T.H.M. was supported by grants from the Novo Nordisk Foundation (NNF20OC0064890 and NNF21OC0067157). C.M.B. is supported by a

MSFHR Health Professional-Investigator Award. P.Q.H. and L.H. were funded by the European Union's Horizon 2020 research and innovation program (ATAC, 101003650). Work at Y.-L.L.'s laboratory in the University of Hong Kong (HKU) was supported by the Society for the Relief of Disabled Children. MBBS/PhD study of D.L. in HKU was supported by the Croucher Foundation. J.L.F. was supported in part by the Coopération Scientifique France-Colciencias (ECOS-Nord/COLCIENCIAS/MEN/ICETEX (806-2018) and Colciencias contract 713-2016 (code 111574455633)]. A.K. was in part supported by grants NU20-05-00282 and NV18-05-00162 issued by the Czech Health Research Council and Ministry of Health, Czech Republic. L.P. was funded by Program Project COVID-19 OSR-UniSR and Ministero della Salute (COVID-2020-12371617). I.M. is a Senior Clinical Investigator at the Research Foundation–Flanders and is supported by the CSL Behring Chair of Primary Immunodeficiencies; by the KU Leuven C1 grant C16/18/007; by a VIB-GC PID grant; by the FWO grants G0C8517N, G0B5120N, and G0E8420N; and by the Jeffrey Modell Foundation. I.M. has received funding under the European Union's Horizon 2020 research and innovation program (grant agreement no. 948959). E.A. received funding from the Hellenic Foundation for Research and Innovation (INTERFLU, no. 1574). M.Vi received funding from the São Paulo Research Foundation (FAPESP) (grant number 2020/09702-1) and JBS SA (grant number 69004). The NH-COVAIR study group consortium was supported by a grant from the Meath Foundation.

## **Conflict of Interest Disclosures**

J.-L.C. is an inventor on patent application PCT/US2021/042741, filed 22 July 2021, submitted by The Rockefeller University, which covers diagnosis of, susceptibility to, and treatment of viral disease and viral vaccines, including COVID-19 and vaccine-associated diseases. M.C.N. is an inventor on patent application PCT/US2021/070472 submitted by The Rockefeller University that covers neutralizing anti-SARS-CoV-2 antibodies and methods of the use thereof. M.C.N. reports being on the Scientific Advisory Board of Celldex and Frontier Biotechnologies. R.P.L. reports being a non-executive director of Roche.

## **Availability of data and materials**

All the data are available in the manuscript or in the Supplementary Materials. Plasma, cells, and genomic DNA are available from J.-L.C. under a material transfer agreement (MTA) with The Rockefeller University or the Imagine Institute. Huh-7.5 cells are available on request from C.M.R. under an MTA with The Rockefeller University and Apath LLC. The materials and reagents used are almost exclusively commercially available and nonproprietary. Materials derived from human samples may be made available on request, subject to any underlying restrictions concerning such samples.

# Code availability

Access to the code is available from the authors on request for noncommercial, academic and research use only.

## Group Information

Lists of members of the HGID Lab, COVID Clinicians, COVID-STORM Clinicians, NIAID Immune Response to COVID Group, NH-COVAIR Study Group, Danish CHGE, Danish Blood Donor Study, St. James's Hospital, SARS CoV2 Interest group, French COVID Cohort Study Group, Imagine COVID-Group, The Milieu Intérieur Consortium, CoV-Contact Cohort, Amsterdam UMC Covid-19 Biobank Investigators, COVID Human Genetic Effort, CP-COVID-19 Group, CONSTANCES cohort, 3C-Dijon Study, Cerba Health-Care, Etablissement du Sang study group consortia are available in the Supplementary Information.

## References

1. Hu, B., Guo, H., Zhou, P. & Shi, Z.L. Characteristics of SARS-CoV-2 and COVID-19. *Nat Rev Microbiol* **19**, 141-154 (2021).
2. Pei, S., Yamana, T.K., Kandula, S., Galanti, M. & Shaman, J. Burden and characteristics of COVID-19 in the United States during 2020. *Nature* **598**, 338-341 (2021).
3. Zhang, Q., *et al.* Life-Threatening COVID-19: Defective Interferons Unleash Excessive Inflammation. *Med (N Y)* **1**, 14-20 (2020).
4. O'Driscoll, M., *et al.* Age-specific mortality and immunity patterns of SARS-CoV-2. *Nature* **590**, 140-145 (2021).
5. Levin, A.T., *et al.* Assessing the age specificity of infection fatality rates for COVID-19: systematic review, meta-analysis, and public policy implications. *Eur J Epidemiol* **35**, 1123-1138 (2020).
6. Brodin, P. Immune determinants of COVID-19 disease presentation and severity. *Nat Med* **27**, 28-33 (2021).
7. Casanova, J.L., Su, H.C. & Effort, C.H.G. A Global Effort to Define the Human Genetics of Protective Immunity to SARS-CoV-2 Infection. *Cell* **181**, 1194-1199 (2020).
8. Bastard, P., *et al.* Autoantibodies neutralizing type I IFNs are present in ~4% of uninfected individuals over 70 years old and account for ~20% of COVID-19 deaths. *Sci Immunol* **6**(2021).

9. Bastard, P., *et al.* Autoantibodies against type I IFNs in patients with life-threatening COVID-19. *Science* **370**(2020).
10. Asano, T., *et al.* X-linked recessive TLR7 deficiency in ~1% of men under 60 years old with life-threatening COVID-19. *Sci Immunol* **6**(2021).
11. Zhang, Q., *et al.* Inborn errors of type I IFN immunity in patients with life-threatening COVID-19. *Science* **370**(2020).
12. Khanmohammadi, S., Rezaei, N., Khazaei, M. & Shirvani, A. A Case of Autosomal Recessive Interferon Alpha/Beta Receptor Alpha Chain (IFNAR1) Deficiency with Severe COVID-19. *J Clin Immunol* (2021).
13. Schmidt, A., *et al.* TBK1 and TNFRSF13B mutations and an autoinflammatory disease in a child with lethal COVID-19. *NPJ Genom Med* **6**, 55 (2021).
14. Abolhassani, H., *et al.* X-Linked TLR7 Deficiency Underlies Critical COVID-19 Pneumonia in a Male Patient with Ataxia-Telangiectasia. *J Clin Immunol* (2021).
15. Casanova, J.L. & Abel, L. Mechanisms of viral inflammation and disease in humans. *Science* **374**, 1080-1086 (2021).
16. Goncalves, D., *et al.* Antibodies against type I interferon: detection and association with severe clinical outcome in COVID-19 patients. *Clin Transl Immunology* **10**, e1327 (2021).
17. Koning, R., *et al.* Autoantibodies against type I interferons are associated with multi-organ failure in COVID-19 patients. *Intensive Care Med* **47**, 704-706 (2021).
18. Troya, J., *et al.* Neutralizing Autoantibodies to Type I IFNs in >10% of Patients with Severe COVID-19 Pneumonia Hospitalized in Madrid, Spain. *J Clin Immunol* **41**, 914-922 (2021).
19. van der Wijst, M.G.P., *et al.* Type I interferon autoantibodies are associated with systemic immune alterations in patients with COVID-19. *Sci Transl Med* **13**, eabh2624 (2021).
20. Vazquez, S.E., *et al.* Neutralizing Autoantibodies to Type I Interferons in COVID-19 Convalescent Donor Plasma. *J Clin Immunol* **41**, 1169-1171 (2021).
21. Wang, E.Y., *et al.* Diverse functional autoantibodies in patients with COVID-19. *Nature* **595**, 283-288 (2021).
22. Abers, M.S., *et al.* Neutralizing type-I interferon autoantibodies are associated with delayed viral clearance and intensive care unit admission in patients with COVID-19. *Immunol Cell Biol* **99**, 917-921 (2021).

23. Chauvineau-Grenier, A., *et al.* Autoantibodies neutralizing type I interferons in 20% of COVID-19 deaths in a French hospital. *Res Sq* (2021).
24. Solanich, X., *et al.* Pre-existing Autoantibodies Neutralizing High Concentrations of Type I Interferons in Almost 10% of COVID-19 Patients Admitted to Intensive Care in Barcelona. *J Clin Immunol* (2021).
25. Raadsen, M.P., *et al.* Interferon- $\alpha$ 2 Auto-antibodies in Convalescent Plasma Therapy for COVID-19. *Journal of Clinical Immunology* (2021).
26. Chang, S.E., *et al.* New-onset IgG autoantibodies in hospitalized patients with COVID-19. *Nat Commun* **12**, 5417 (2021).
27. Ziegler, C.G.K., *et al.* Impaired local intrinsic immunity to SARS-CoV-2 infection in severe COVID-19. *Cell* **184**, 4713-4733 e4722 (2021).
28. Acosta-Ampudia, Y., *et al.* COVID-19 convalescent plasma composition and immunological effects in severe patients. *J Autoimmun* **118**, 102598 (2021).
29. Carapito, R., *et al.* Identification of driver genes for critical forms of COVID-19 in a deeply phenotyped young patient cohort. *Sci Transl Med*, eabj7521 (2021).
30. Bastard, P., *et al.* Preexisting autoantibodies to type I IFNs underlie critical COVID-19 pneumonia in patients with APS-1. *J Exp Med* **218**(2021).
31. van der Wijst, M.G.P., *et al.* Type I interferon autoantibodies are associated with systemic immune alterations in patients with COVID-19. *Sci Transl Med* (2021).
32. Galani, I.E., *et al.* Untuned antiviral immunity in COVID-19 revealed by temporal type I/III interferon patterns and flu comparison. *Nat Immunol* **22**, 32-40 (2021).
33. Sposito, B., *et al.* The interferon landscape along the respiratory tract impacts the severity of COVID-19. *Cell* **184**, 4953-4968 e4916 (2021).
34. Lopez, J., *et al.* Early nasal type I IFN immunity against SARS-CoV-2 is compromised in patients with autoantibodies against type I IFNs. *J Exp Med* **218**(2021).
35. Blanco-Melo, D., *et al.* Imbalanced Host Response to SARS-CoV-2 Drives Development of COVID-19. *Cell* **181**, 1036-1045 e1039 (2020).
36. Israelow, B., *et al.* Mouse model of SARS-CoV-2 reveals inflammatory role of type I interferon signaling. *J Exp Med* **217**(2020).
37. Doffinger, R., *et al.* Autoantibodies to interferon-gamma in a patient with selective susceptibility to mycobacterial infection and organ-specific autoimmunity. *Clin Infect Dis* **38**, e10-14 (2004).

38. Hoflich, C., *et al.* Naturally occurring anti-IFN-gamma autoantibody and severe infections with *Mycobacterium chelonae* and *Burkholderia cocovenenans*. *Blood* **103**, 673-675 (2004).
39. Kampmann, B., *et al.* Acquired predisposition to mycobacterial disease due to autoantibodies to IFN-gamma. *J Clin Invest* **115**, 2480-2488 (2005).
40. Puel, A., *et al.* Recurrent staphylococcal cellulitis and subcutaneous abscesses in a child with autoantibodies against IL-6. *J Immunol* **180**, 647-654 (2008).
41. Puel, A., *et al.* Autoantibodies against IL-17A, IL-17F, and IL-22 in patients with chronic mucocutaneous candidiasis and autoimmune polyendocrine syndrome type I. *J Exp Med* **207**, 291-297 (2010).
42. Ku, C.L., Chi, C.Y., von Bernuth, H. & Doffinger, R. Autoantibodies against cytokines: phenocopies of primary immunodeficiencies? *Hum Genet* **139**, 783-794 (2020).
43. Kisand, K., *et al.* Chronic mucocutaneous candidiasis in APECED or thymoma patients correlates with autoimmunity to Th17-associated cytokines. *J Exp Med* **207**, 299-308 (2010).
44. Rosen, L.B., *et al.* Nocardia-induced granulocyte macrophage colony-stimulating factor is neutralized by autoantibodies in disseminated/extrapulmonary nocardiosis. *Clin Infect Dis* **60**, 1017-1025 (2015).
45. Casanova, J.L. & Abel, L. Lethal Infectious Diseases as Inborn Errors of Immunity: Toward a Synthesis of the Germ and Genetic Theories. *Annu Rev Pathol* **16**, 23-50 (2021).
46. Bastard, P., *et al.* Auto-antibodies to type I IFNs can underlie adverse reactions to yellow fever live attenuated vaccine. *J Exp Med* **218**(2021).
47. Morabia, A., Ten Have, T. & Landis, J.R. Empirical evaluation of the influence of control selection schemes on relative risk estimation: the Welsh nickel workers study. *Occup Environ Med* **52**, 489-493 (1995).
48. Bartleson, J.M., *et al.* SARS-CoV-2, COVID-19 and the Ageing Immune System. *Nat Aging* **1**, 769-782 (2021).
49. Splunter, M.V., *et al.* Plasmacytoid dendritic cell and myeloid dendritic cell function in ageing: A comparison between elderly and young adult women. *PLoS One* **14**, e0225825 (2019).
50. Schultze, J.L. & Aschenbrenner, A.C. COVID-19 and the human innate immune system. *Cell* **184**, 1671-1692 (2021).
51. Stark, G.R. & Darnell, J.E., Jr. The JAK-STAT pathway at twenty. *Immunity* **36**, 503-514 (2012).
52. Pierce, C.A., *et al.* Natural mucosal barriers and COVID-19 in children. *JCI Insight* **6**(2021).



53. Manry, J., *et al.* Evolutionary genetic dissection of human interferons. *J Exp Med* **208**, 2747-2759 (2011).
54. Ng, C.T., *et al.* Blockade of interferon Beta, but not interferon alpha, signaling controls persistent viral infection. *Cell Host Microbe* **17**, 653-661 (2015).
55. Vinh, D.C., *et al.* Harnessing Type I IFN Immunity Against SARS-CoV-2 with Early Administration of IFN-beta. *J Clin Immunol* **41**, 1425-1442 (2021).
56. Weinreich, D.M., *et al.* REGN-COV2, a Neutralizing Antibody Cocktail, in Outpatients with Covid-19. *N Engl J Med* **384**, 238-251 (2021).
57. Bastard, P., *et al.* Interferon-beta Therapy in a Patient with Incontinentia Pigmenti and Autoantibodies against Type I IFNs Infected with SARS-CoV-2. *J Clin Immunol* **41**, 931-933 (2021).
58. Monk, P.D., *et al.* Safety and efficacy of inhaled nebulised interferon beta-1a (SNG001) for treatment of SARS-CoV-2 infection: a randomised, double-blind, placebo-controlled, phase 2 trial. *Lancet Respir Med* **9**, 196-206 (2021).

## Tables

**Table 1. Lines of evidence suggesting that auto-Abs against type I IFNs are strong determinants of the risk of life-threatening COVID-19.**

Evidence	Examples	References
Auto-Abs against type I IFNs are present before SARS-CoV-2 infection	In patients for whom a sample collected before the COVID-19 pandemic was available, the auto-Abs were found to pre-exist infection	30
	These auto-Abs are found in the uninfected general population, and their prevalence increases after the age of 65 years	8
Auto-Abs are associated with COVID-19 severity	Patients with inborn errors underlying these auto-Abs from infancy onward (e.g. APS-1) have a very high risk of developing critical COVID-19 pneumonia	30
	The population of patients with critical disease includes a higher proportion of individuals producing these auto-Abs than the population of patients with silent or mild infection (ORs depending on the nature, number, and concentrations of type I IFN neutralized)	8
	The results concerning the proportions of critical cases with auto-Abs against type I IFNs have already been replicated in >15 different cities (Americas, Europe, Asia)	16-29
Auto-Abs against type I IFNs neutralize host antiviral activity	These auto-Abs neutralize the antiviral activity of type I IFNs against SARS-CoV-2 <i>in vitro</i>	9
	These auto-Abs are found <i>in vivo</i> in the blood of SARS-CoV-2-infected patients, where they neutralize type I IFN	31
	These auto-Abs are found <i>in vivo</i> in the respiratory tract of patients, where they neutralize type I IFN	32-34
	A key virulence factor of SARS-CoV-2 <i>in vitro</i> is its capacity to impair type I IFN immunity	35
	Animals with type I IFN deficiency develop critical disease, including animals treated with mAbs that neutralize type I IFNs	36
Auto-Abs against cytokines are clinical phenocopies of the corresponding inborn errors	Patients with auto-Abs against type I IFNs are phenocopies of IFNAR1 <sup>-/-</sup> , IFNAR2 <sup>-/-</sup> , and IRF7 <sup>-/-</sup> patients with critical COVID-19 pneumonia	11
	Patients with auto-Abs against IL-6, IL-17, GM-CSF, and type II IFN are phenocopies of the corresponding inborn errors and underlie staphylococcal disease, mucocutaneous candidiasis, nocardiosis, and mycobacterial diseases, respectively	37-45

**Table 2. Characteristics of the general population cohort and of the cohort of patients who died from COVID-19, by age, sex and autoantibody status**

Characteristics	Neutralization 100 pg/mL		Neutralization 10 ng/mL	
	General Population (N=10,778)	Deceased Patients (N=1,121)	General Population (N=34,159)	Deceased Patients (N=1,094)
Male - no. (%)	5,429 (50.4) <sup>a</sup>	821 (73.2)	17,859 (52.3)	805 (73.5)
Mean age <sup>a</sup> ±SD - yr	62.3 ±17.2	70.7 ±13.0	52.7 ±18.2	70.6 ±13.1
Age distribution - no. (%)				
20-39 yr	1,251 (11.6)	17 (1.5)	9,102 (26.6)	15 (1.4)
40-49 yr	1,459 (13.5)	43 (3.8)	5,403 (15.8)	47 (4.3)
50-59 yr	1,736 (16.1)	144 (12.8)	6,414 (18.9)	152 (13.9)
60-69 yr	2,475 (23.0)	307 (27.4)	6,881 (20.1)	289 (26.4)
70-79 yr	1,790 (16.6)	307 (27.4)	3,721 (10.9)	296 (27.1)
≥80 yr	2,067 (19.2)	303 (27.0)	2,638 (7.7)	295 (27.0)
Auto-Ab - no. of carriers (%)				
IFN-α2 and IFN-ω	65 (0.6)	102 (9.1)	45 (0.1)	75 (6.8)
IFN-α2 or IFN-ω	246 (2.3)	203 (18.1)	181 (0.5)	130 (11.9)
IFN-α2	151 (1.4)	140 (12.5)	117 (0.3)	118 (10.8)
IFN-ω	160 (1.5)	165 (14.7)	109 (0.3)	87 (8.0)
IFN-β <sup>b</sup>	NA	NA	24 (0.3)	6 (0.9)

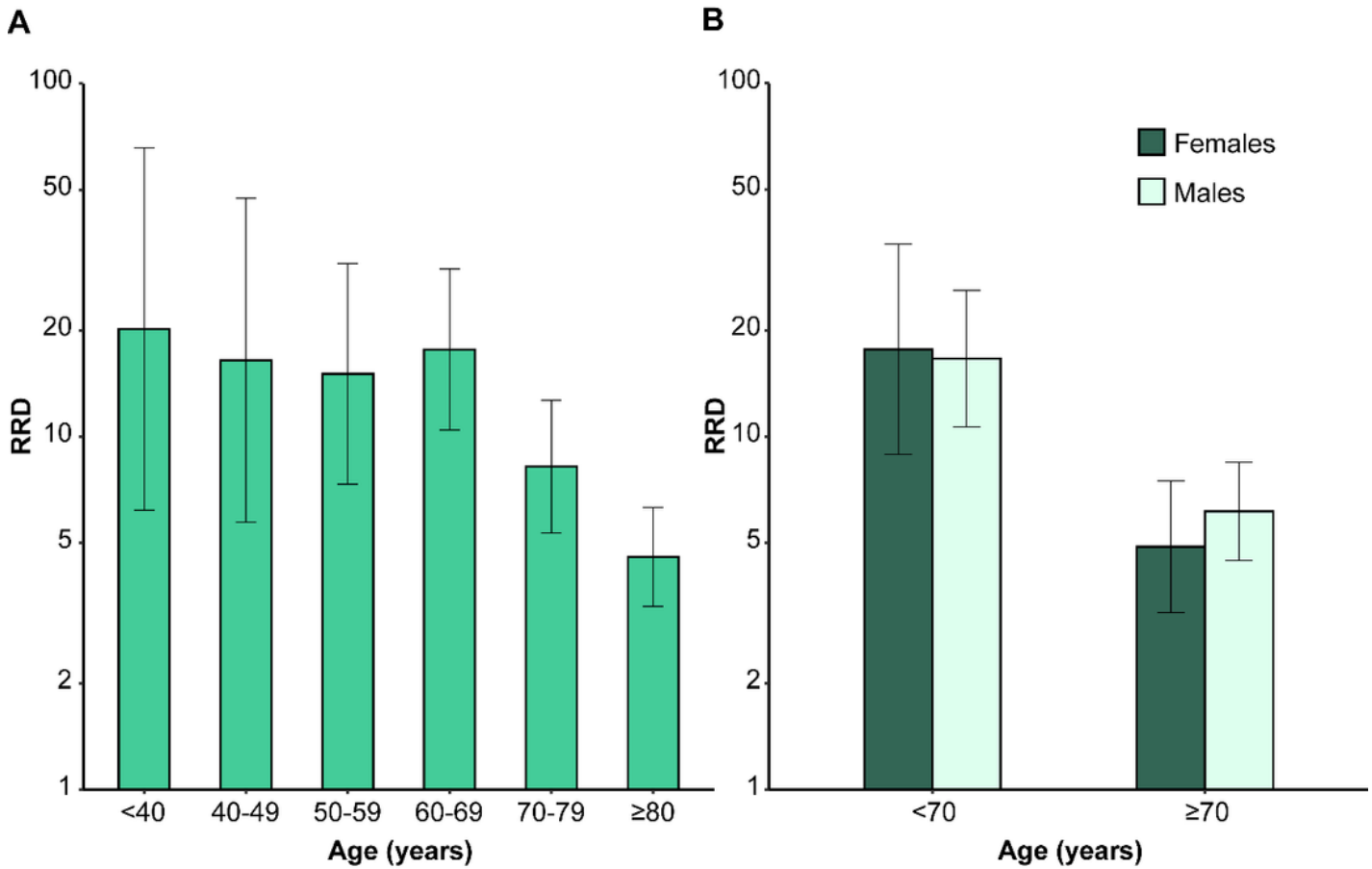
SD standard deviation.

NA not available.

<sup>a</sup>Age is given in years and corresponds to age at the time of recruitment for members of the general population cohort (controls) and age at death for COVID-19 patients.

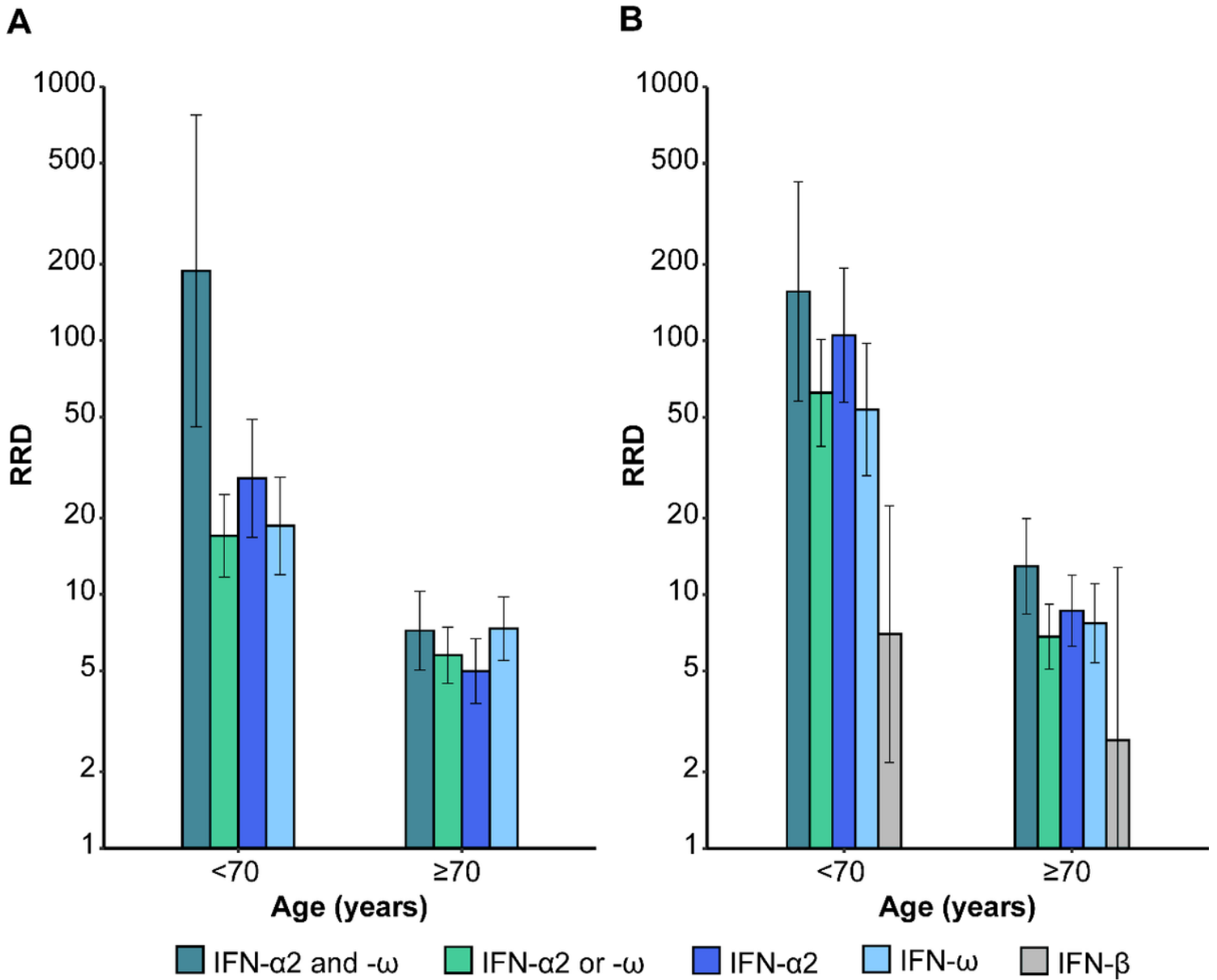
<sup>b</sup>IFN-β neutralization experiments were performed only for a concentration of 10 ng/mL, on 9,126 individuals (49.5% male, mean age 60.6 years) from the general population and 636 COVID-19 patients (71.1% male, mean age 72.9 years).

## Figures



**Figure 1**

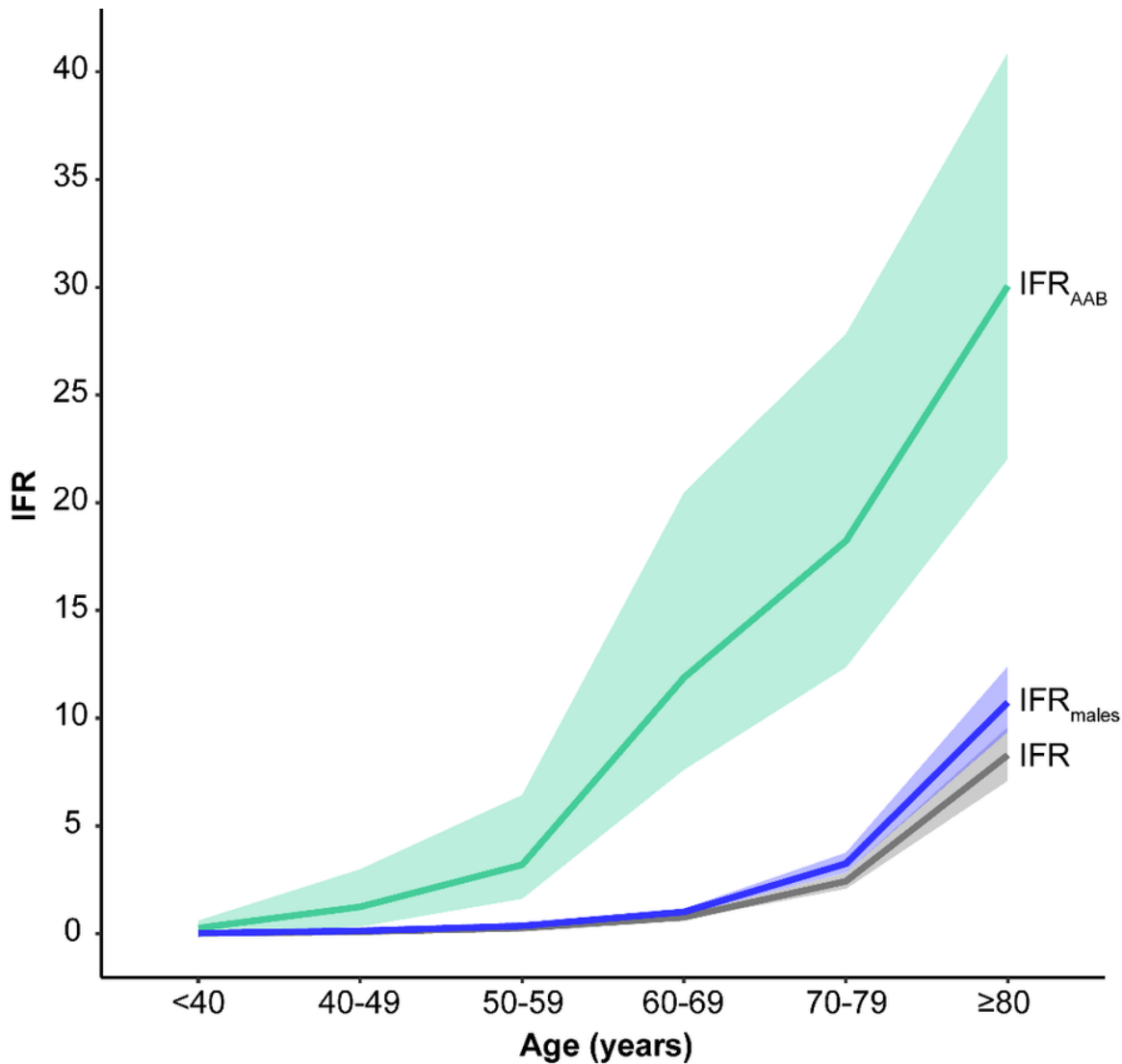
**Relative risks of death associated with auto-Abs neutralizing low concentrations of IFN- $\alpha$ 2 or - $\omega$ , by age and sex.** RRDs for individuals with auto-Abs neutralizing low concentrations of IFN- $\alpha$ 2 or IFN- $\omega$  relative to individuals without such auto-Abs, by age and sex. RRDs are displayed on a logarithmic scale (A) for six age classes, and (B) for male and female subjects under and over the age of 70 years. Vertical bars represent the 95% CI.



**Figure 2**

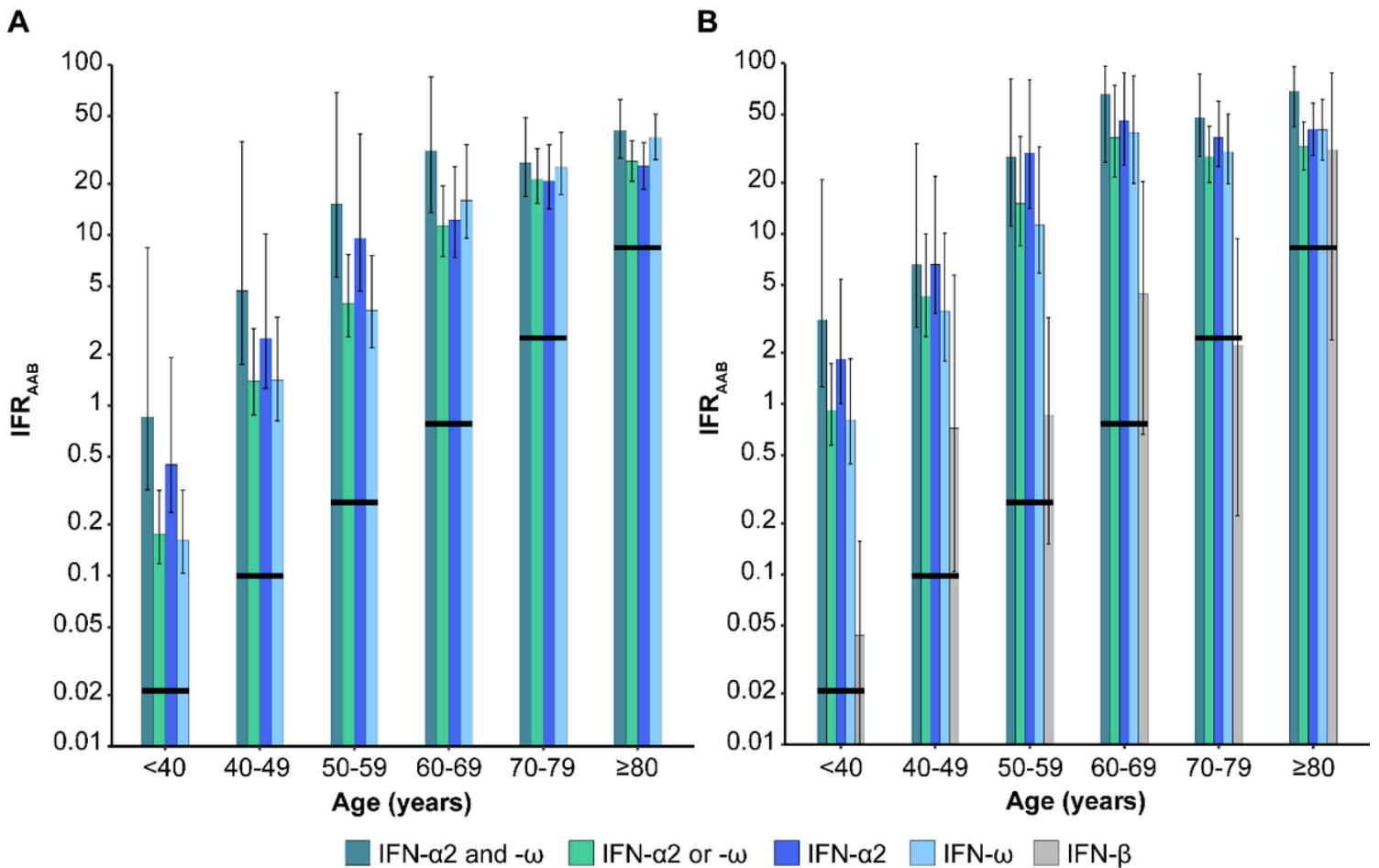
**Relative risks of death associated with auto-Abs neutralizing various combinations of type I IFNs, by age.**

RRDs for individuals with auto-Abs neutralizing different combinations of type I IFNs relative to individuals without such auto-Abs, by age. RRDs are displayed on a logarithmic scale for individuals under and over 70 years of age with (A) auto-Abs neutralizing low concentrations of IFN-α2 and IFN-ω, IFN-α2 or IFN-ω, IFN-α2, IFN-ω, and (B) auto-Abs neutralizing high concentrations of IFN-α2 and IFN-ω, IFN-α2, IFN-ω and IFN-β, relative to individuals without such combinations of auto-Abs. Vertical bars represent the 95% CI.



**Figure 3**

**SARS-CoV-2 infection fatality rates by age.** IFRs are provided in the general population for both sexes (gray) and for males only (blue) using the data of O’Driscoll et al. <sup>4</sup>; IFR<sub>AAB</sub> (green) are shown for individuals carriers of auto-Abs neutralizing low concentrations of IFN- $\alpha$ 2 or IFN- $\omega$ . Auto-Abs against type I IFNs are associated with high RRDs and strongly increase IFR, to a much greater extent than maleness, and by inference than other classical common risk factors providing ORs of death similar to maleness (around 2) such as some comorbidities, or the most significant common genetic variant on chromosome 3<sup>3</sup>.



**Figure 4**

**SARS-CoV-2 infection fatality rates for carriers of various combinations of neutralizing auto-Abs, by age.**

IFR<sub>AAB</sub> values (%) are displayed, on a logarithmic scale, by age, for individuals with (A) auto-Abs neutralizing low concentrations of IFN-α2 and IFN-ω, IFN-α2 or IFN-ω, IFN-α2, IFN-ω, and (B) auto-Abs neutralizing high concentrations of IFN-α2 and IFN-ω, IFN-α2 or IFN-ω, IFN-α2, IFN-ω and IFN-β. Vertical bars represent the 95% CI. Horizontal black lines represent the IFR provided by O'Driscoll *et al*<sup>4</sup>.

**Supplementary Files**

This is a list of supplementary files associated with this preprint. Click to download.

- [SupplementaryInformation20220103.docx](#)